


Review

# The Role of Inflammation and Inflammatory Mediators in the Development, Progression, Metastasis, and Chemoresistance of Epithelial Ovarian Cancer

Sudha S. Savant <sup>1,†</sup>, Shruthi Sriramkumar <sup>2,†</sup> and Heather M. O'Hagan <sup>1,3,\*</sup> 

<sup>1</sup> Medical Sciences, Indiana University School of Medicine, Bloomington, IN 47405, USA; ssavant@iu.edu

<sup>2</sup> Cell, Molecular and Cancer Biology Graduate Program, Indiana University, Bloomington, IN 47405, USA; ssriramk@iu.edu

<sup>3</sup> Indiana University Melvin and Bren Simon Cancer Center, Indianapolis, IN 46202, USA

\* Correspondence: hmohagan@indiana.edu; Tel.: +1-812-855-3035

† These authors contributed equally to this work.

Received: 14 June 2018; Accepted: 24 July 2018; Published: 30 July 2018



**Abstract:** Inflammation plays a role in the initiation and development of many types of cancers, including epithelial ovarian cancer (EOC) and high grade serous ovarian cancer (HGSC), a type of EOC. There are connections between EOC and both peritoneal and ovulation-induced inflammation. Additionally, EOCs have an inflammatory component that contributes to their progression. At sites of inflammation, epithelial cells are exposed to increased levels of inflammatory mediators such as reactive oxygen species, cytokines, prostaglandins, and growth factors that contribute to increased cell division, and genetic and epigenetic changes. These exposure-induced changes promote excessive cell proliferation, increased survival, malignant transformation, and cancer development. Furthermore, the pro-inflammatory tumor microenvironment environment (TME) contributes to EOC metastasis and chemoresistance. In this review we will discuss the roles inflammation and inflammatory mediators play in the development, progression, metastasis, and chemoresistance of EOC.

**Keywords:** inflammation; epithelial ovarian cancer; cytokines; reactive oxygen species; growth factors

## 1. Inflammation and EOC

Inflammation is part of the immune response that protects against foreign pathogens and aids in healing. Inflammation is elicited in response to cellular damage either by infection, exposure to foreign particles (pollutants or irritants), or an increase in cellular stress [1]. The ultimate goal of the inflammatory response is to restore tissue homeostasis, either by destruction or healing of the damaged tissue. The acute or immediate inflammatory response involves modification of the vasculature surrounding the site of stress or damage to increase blood flow. This alteration is then followed by activation of innate immune cells already present in the tissue, including macrophages, dendritic cells (DC), and mast cells, and an increase in infiltration of additional innate immune cells into the affected tissue. At sites of inflammation there are high levels of reactive oxygen species (ROS), cytokines, chemokines, and growth factors that are produced by the immune cells and other cells in the tissue. Acute inflammation is essential for tissue homeostasis and to protect against normal exposure to pathogens. However, in certain cases the body is unable to resolve this response or is subjected to repeated stimulation resulting in chronic inflammation.

Ovarian cancer (OC) is the fifth leading cause of cancer-related deaths in women in the United States [2] and can originate in the germ cells, sex-cord stroma, the fallopian tube (FT), or ovary

epithelium. Epithelial ovarian cancer (EOC) which originates from the ovary or fallopian tube epithelium, accounts for 85–90% of all OCs. Chronic inflammation is an important risk factor associated for EOC and high grade serous ovarian cancer (HGSC), the most malignant subtype of EOC. Chronic inflammation results in activation of signaling pathways, transcription factors, and the innate and adaptive immune responses [3,4]. In this review we primarily focus on inflammation as a risk factor for invasive EOC, but have also included supportive evidence from other OC subtypes, studies that do not define the subtype of OC, and other tumor types as indicated.

### 1.1. Signaling Pathways and Transcription Factors

Several signaling pathways and transcription factors involved in the inflammatory response also play critical roles in EOC. Here we briefly introduce relevant pathways that will be linked to OC formation in later sections. Cytokines produced during inflammation bind to and activate toll like receptors (TLRs) on cell surfaces, which results in activation of the signaling pathways involving mitogen-activated protein kinases (MAPKs) p38 and JNK (c-Jun N-terminal kinase) and transcription factors including nuclear factor kappa-light-chain-enhancer of activated B cells (NF- $\kappa$ B) and the signal transducer and activator of transcription (STATs). The MAPK pathway regulates cellular processes like proliferation, differentiation, growth, migration, and cell death by upregulating the expression of transcription factors like AP-1, c-Jun, FOS and by activating NF- $\kappa$ B and STATs, that either by themselves or along with AP-1 or c-Jun regulate expression of pro-survival and pro-growth genes. NF- $\kappa$ B and AP-1 also regulate production of cytokines like IL-6 [5–7].

During inflammation these transcription factors play an important role to maintain tissue homeostasis. However, in case of chronic inflammation, the signaling pathways are continuously stimulated, which can contribute to tumorigenesis.

### 1.2. Innate Immune Response

Inflammation activates the innate immune response, which signals macrophages and DCs to secrete chemoattractants like Interleukin-8 (IL-8), monocyte chemotactic protein-1 (MCP-1), and various other inflammatory mediators. These chemoattractants in turn result in recruitment of neutrophils, lymphocytes, and natural killer (NK) cells to the site of damage. All of these cells then secrete cytokines like IL-1, IL-3, IL-6, IL-8, tumor necrosis factor alpha (TNF- $\alpha$ ), interferon (IFN)  $\alpha$ , and colony-stimulating factors (CSF) like granulocyte macrophage CSF (GM-CSF). The cytokines bind to transmembrane receptors on the cell surfaces of other cells to activate transcription factors that regulate gene expression downstream of the cytokine activated pathway. This creates a pro-inflammatory environment resulting in recruitment of other immune cells, migration of endothelial cells, and proliferation of fibroblasts. Activation of macrophages and NK cells results in the production of high levels of ROS and reactive nitrogen species (RNS), which are used by these cells to kill foreign pathogens, but also end up damaging neighboring normal cells [8]. The lymphocytes also secrete growth factors like platelet derived growth factor (PDGF), transforming growth factor beta (TGF- $\beta$ ), and fibroblast growth factor (FGF), which facilitate wound healing. Overall the acute immune response is a rapid response that typically only lasts a few days. It results in removal of the pathogen, release of proteolytic enzymes to destroy damaged tissue, or stimulation of the proliferation of fibroblasts and epithelial cells to repair the tissue [1].

### 1.3. Adaptive Immune Response

If the infection is not resolved by the innate immune response, the adaptive immune response is activated, which is less inflammatory in nature. The adaptive immune response also provides longstanding protection against specific pathogens and/or antigens. B cells and T cells are the effector cells of the adaptive immune system that are derived from lymphocytes when they are presented with specific antigens by the antigen presenting cells (APC). T cells respond to the APCs by producing IL-2, which induces expression of transcription factors that facilitate T cells to differentiate

into T regulatory (Tregs) and T effector (Teff) cells. There are two major classes of T effector cells; CD8<sup>+</sup> cytotoxic T cells and CD4<sup>+</sup> T helper (Th) cells. Th cells are further differentiated into Th1, Th2, or Th17 depending on the ILs secreted and the transcription factors expressed. IFN- $\gamma$  activates STAT1 to induce formation of Th1 and IL-6, and TGF- $\beta$  can induce Th17 cell formation. Th1 and Th17 secrete ILs and activate macrophages and B cells to create a pro-inflammatory microenvironment (ME) that can be protumorigenic depending on the context. Tregs are immunosuppressive cells that turn off the immune response [1,9,10].

## 2. Inflammation as a Risk Factor for EOC

Amongst other factors such as hereditary, environmental, and lifestyle, inflammation emerges as an important risk factor for EOC. EOC arises either in the epithelial layer surrounding the ovary or in the epithelium of the distal FT, which could then spread to the ovary. A significant portion of HGSC is thought to originate in the FT, in part because removal of the FT significantly reduces OC risk [11]. Interestingly, while surgical specimens from mutation carriers rarely had premalignant ovarian epithelial changes, early lesions called serous tubal intraepithelial carcinomas (STICs) were found in the FTs of 5–10% of the patients. Copy number and mutational analysis suggest that STICs shed cells with metastatic potential that then colonize the ovary to form HGSC. STICs are mostly found in the fimbriae, the distal end of the FT that shares a ME with the ovary. During a woman's lifetime, the repeated secretion of ROS, cytokines, and other growth factors by the ovaries and immune cells creates a chronic inflammatory ME in the peritoneum that in turn potentiates the initiation of normal cells to malignant ones in the FT and the ovary, supports tumor progression, metastasis, and development of resistance to chemotherapy.

During ovulation, infection and other causes of inflammation ovary and FT tissue is damaged and undergoes repair. We will briefly discuss how each of these processes evoke or involve an inflammatory response that can persist, leading to a cytokine and growth factor rich environment in the peritoneum and contribute to EOC.

### 2.1. Ovulation

The process of ovulation itself is comparable to that of inflammation as described in the early 20th century. The development of the follicle to its rupture and release of the egg results in recruitment of activated immune cells to the ovary and production of enormous amounts of chemokines, cytokines, and growth factors. Ovulation is initiated by a surge of Luteinizing hormones (LH) that results in increased blood flow to the ovarian follicles. Before release of the egg, the surge of LH hormone recruits neutrophils and macrophages to the graafian follicles [12–14]. Macrophages in the theca have been shown to support growth of follicles [15]. During ovulation macrophages secrete growth factors like hepatocyte growth factor (HGF), TGF- $\beta$ , and epidermal growth factor (EGF), which stimulate cellular proliferation and follicle growth. Simultaneously the macrophages also secrete ROS, TNF- $\alpha$ , and IL1 $\beta$ , which stimulate local apoptosis resulting in rupture of the follicle, which bathes the ovarian surface and fimbriae with follicular fluid. Exposure of FT cells to follicular fluid results in altered expression of genes associated with inflammation, including increased expression of IL8 and cyclooxygenase-2 (COX-2) [16]. Quiescent fibroblasts are present in the thecal layer surrounding the follicles. Exposure to growth factors stimulates their proliferation and they then secrete prostaglandins, collagenases, and plasminogen activator. In the corpus luteum, after the follicle is released, the macrophages secrete prostaglandins, ROS, and TNF- $\alpha$ , which stimulate apoptosis of the corpus luteum cells. Therefore, ovulation results in the cyclic exposure of FT and ovarian epithelial cells to high levels of ROS, cytokines, and growth factors [17] Although the other causes of inflammation discussed below are important and result in increased overall risk for EOC, the process of ovulation itself occurs often in the lifetime of the majority of women and may be the most important inflammation-related risk factor for EOC. This hypothesis is corroborated by the laying hen model, which is commonly used to study ovarian cancer [18]. In this model, hens develop spontaneous EOC, likely due to their high ovulation

rate, thus linking ovulation directly as an increased risk factor for EOC. Delayed onset of menarche and early onset of menopause have been shown to be inversely related to the risk of OC, likely due to the reduction in number of ovulation cycles in a woman's lifetime [19,20]. Further, ovulation has also been connected to EOC because contraceptive pills, pregnancy, and breastfeeding reduce the risk of OC. These factors reduce, halt, or delay overall ovulation cycles, respectively, which in turn reduces overall exposure to inflammation of the ovary and FT. The associations of parity and oral contraceptive use with invasive EOC were recently confirmed in a large, prospective study using the European Prospective Investigation into Cancer and Nutrition (EPIC) cohort that found only limited heterogeneity in the risk between reproductive factors and EOC subtypes [21]. Hysterectomy, tube ligation, and removal of ovaries are also protective against development of OC [22,23].

## 2.2. Infection

Pelvic inflammatory disorder (PID) is the infection of the female reproductive organs like cervix, uterus, FTs, and ovaries. It is a significant risk factor for OC and is caused by various bacteria and virus such as *Chlamydia trachomatis*, *Mycoplasma genitalium*, *Neisseria gonorrhoeae*, human papilloma virus, and cytomegalovirus [24,25]. Infection by these microbes results in DNA damage and production of ROS and induces a pro-inflammatory response, which involves secretion of cytokines and migration of immune cells [24]. PID is generally resolved with antibiotics within 48–72 hours of detection. However, repeated infection and unresolved inflammation can lead to chronic inflammation that is a risk factor for EOC.

## 2.3. Other Sources of Inflammation

The other causes of inflammation in the ovaries and/or FTs are endometriosis, obesity, Polycystic Ovarian Syndrome (PCOS), and talc exposure. Endometriosis is defined as presence of stroma and endometrial gland tissues in the pelvic peritoneum, rectovaginal septum, and ovaries [26]. Retrograde menstruation is the most commonly accepted theory for endometriosis. Retrograde menstruation results in aberrant accumulation of red blood cells (RBCs) and tissue, which can trigger an inflammatory response, activating the macrophages in the peritoneal cavity [27,28]. The macrophages lyse the RBCs, resulting in an increase in iron accumulation in the endometrial implants and peritoneal fluid. The accumulated iron can catalyze formation of free radicals like RNS and ROS in the peritoneum and results in increased oxidative stress (OS). OS can activate NF- $\kappa$ B, in macrophages resulting in secretion of growth factors, cytokines, and IFNs. Around one third of women are affected by mild endometriosis, which resolves on its own over time. For the remaining cases, endometriosis results in chronic pain and inflammation, which can be resolved by excision of affected tissue or the outgrowth. However, in 45% of these cases, the endometriosis reoccurs resulting in repeated bouts of chronic inflammation [29,30].

Obese women have higher risks of EOC and HGSC and pro-inflammatory cytokines are associated with higher body mass index (BMI) levels. Adipose tissues secrete the cytokines TNF- $\alpha$ , IL-6, IL-8, and MCP-1, which can induce an inflammatory reaction in the peritoneum [31]. Continuous secretion of these cytokines leads to a state of chronic inflammation, which includes activation of macrophages and recruitment of NK cells and results in high levels of OS. Once the tumor has been initiated, the continuous secretion of cytokines by adipose tissue or omentum can facilitate migration of cancer cells to the omentum, promoting metastasis of the tumor into the peritoneum [30]. High levels (>10 mg/L) of C-reactive protein (CRP), a marker of global inflammation, are associated with an increased risk of EOC [32,33]. IL-6 itself is not a risk factor for EOC but in obese women IL-6 and CRP may be associated with increased EOC risk [33].

PCOS also contributes to inflammation in women and may increase risk of EOC [34]. PCOS is a hormonal disorder occurring in reproductive aged women during which ovaries may develop numerous small collections of fluid and fail to release eggs properly. Obesity, hyperandrogenism, and increased insulin resistance further characterize PCOS. Increased C-Reactive protein (CRP) and

MCP-1 levels, indicative of low-level chronic inflammation, are elevated in women with PCOS [35–38]. Simultaneously chemokines like IL-18, IL-6, and TNF- $\alpha$  are also increased in circulation in women with PCOS [39–42]. The increase in inflammatory mediators correlates positively with BMI, suggesting that increased obesity in women with PCOS may be the source of inflammation. Increased DNA damage and OS is observed in women with PCOS, which may also increase risk for EOC [43]. Evidence linking PCOS directly to EOC is limited due to small study sizes, PCOS being associated with other EOC risk factors such as obesity, and PCOS possibly being only associated with one subtype of EOC, borderline serous [44].

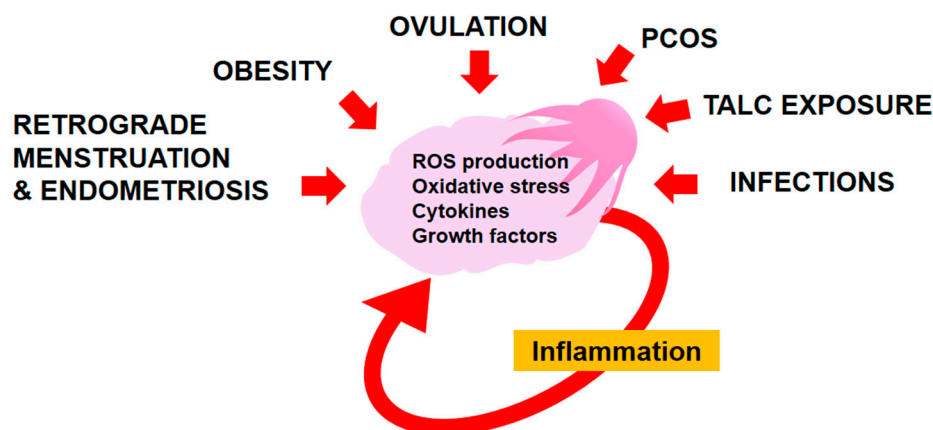
Talc is a silicate mineral and exposure to it can cause inflammation of the ovaries and poses a risk hazard for development of EOC [45]. It has been proposed that talc from talcum powder used for dusting and from condoms and vaginal diaphragms can migrate up to the ovaries via retrograde flow of fluids and mucous and get lodged in the ovaries. Tubal ligation, which is protective for EOC, is thought to block the transport of talc from the lower genital tract. Talc behaves as a foreign particle, triggering an inflammatory response [46,47]. The talc attracts macrophages, which try to phagocytose it. The macrophages then send chemotactic signals to other immune response mediators and initiate a wound healing process. Since talc is not degradable by the body, it inhibits the wound healing process, resulting in chronic inflammation.

#### 2.4. NSAIDs and Reduced Risk of EOC

Further connecting inflammation to EOC are several studies that demonstrate that intake of non-steroidal anti-inflammatory drugs (NSAIDs), specifically of aspirin, correlates inversely with risk of OC and endometrial cancer [48–52]. In vitro studies with OC cell lines and NSAIDs show that NSAIDs and COX-2 inhibitors facilitate apoptosis, however this effect is not dependent on COX-2 and may be due to upregulation of p21, a protein important for cell cycle arrest [53]. Another study by Arango et al., demonstrates that acetylsalicylic acid or aspirin resulted in increased apoptosis via downregulation of Bcl2 in an endometrial cancer cell line [54]. A third study has shown that a selective COX-2 inhibitor, JTE-522, can inhibit proliferation and increase apoptosis of endometrial cancer cells by increasing levels of p53 and p21 and decreasing phosphorylation of retinoblastoma (Rb) protein, which results in its activation; all of which results in cell cycle arrest [55,56]. Simultaneously, there was an increase in caspase-3 activity, which is indicative of increased apoptosis. Another mechanism by which aspirin could facilitate its chemopreventive nature is by inhibiting oxidative induced DNA damage [57]. COX-1 is also expressed in normal ovaries of the laying hen, with expression increasing in post-ovulatory follicles suggesting its importance for or a role in ovulation. With the onset of OC, COX-1 expression is increased [58] and COX-1 inhibition and NSAIDs have shown to decrease proliferation of ascites in the laying hen OC model [59]. Further, when 0.1% aspirin was included in their diet for one year, although the onset of OC was not different, the progression of cancer was slower when compared to hens fed with regular diet [60].

As discussed, inflammation results in secretion of ROS, growth factors, cytokines, and chemokines into the shared environment surrounding the ovary and distal FT. Exposure of normal tissue to these inflammatory mediators results in activation of downstream signaling that can promote the transformation of normal cells or survival of already transformed cells. Once EOC has already formed further exposure of cancer cells to these inflammatory mediators also results in activation of downstream signaling within the cancer cell and in the surrounding tissue, creating an inflammatory environment that can further promote EOC (Figure 1). We will discuss in more detail how key inflammatory mediators contribute to EOC initiation, progression, metastasis, and chemoresistance.



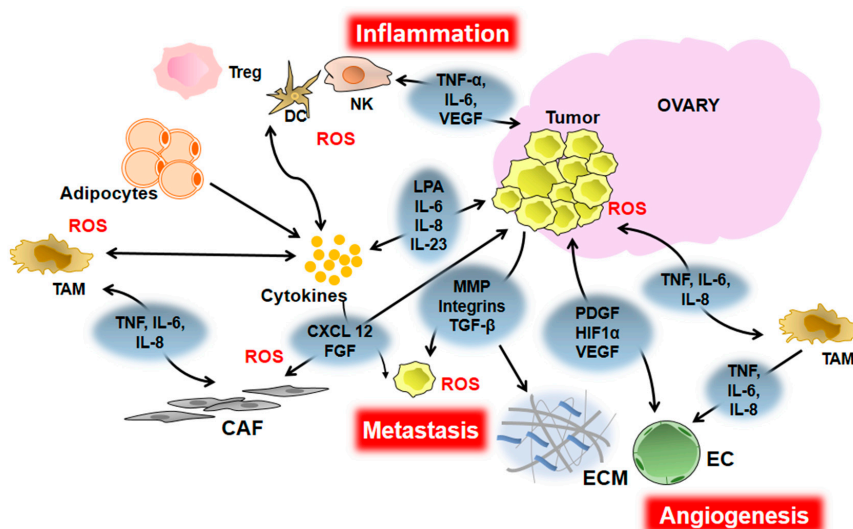


**Figure 1.** Sources of inflammation in the ovary and fimbriae. Ovulation, retrograde menstruation, endometriosis, infections, exposure to talc, Polycystic Ovarian Syndrome (PCOS), and obesity result in exposure of the ovary and fimbriae to reactive oxygen species (ROS), oxidative stress, cytokines, and growth factors, generating an inflammatory response that leads to additional production of ROS and cytokines in the ovary. Unresolved, chronic inflammation is a critical risk factor for tumor initiation.

### 3. Inflammation and EOC Initiation and Progression

Tumorigenesis is a multistep process that requires cells to gain the ability to evade apoptosis and antigrowth signals, proliferate independently of stimuli, develop a support system (angiogenesis), and have the capacity to invade and metastasize. Tumorigenesis is initiated by the transformation of a normal cell to a malignant one. The deregulation of the above mentioned processes in the malignant cell could potentiate its progression to cancer.

One mechanism of cancer initiation is genomic instability due to DNA damage [61] and EOCs exhibit a high number of chromosomal aberrations and genomic instability [62]. The most common gene mutations in HGSCs include *BRCA*, *TP53*, and genes involved in mismatch repair and the DNA damage response [63]. A pro-inflammatory ME can also contribute to genetic instability and therefore play a role in EOC cancer initiation. A pro-inflammatory ME, which is continuously supplemented by ROS, cytokines, and growth factors, can cause DNA damage in epithelial ovarian and FT cells, switch on antiapoptotic pathways, and initiate transformation of normal cells. When cells transformed either by oncogenic alterations or by exposure to inflammation are in a pro-inflammatory ME they are able to turn on pro-survival signaling pathways rather than the senescence pathways that are normally induced by oncogene expression in normal cells. For example, disruption of the RAS pathway results in activated NF- $\kappa$ B signaling and upregulation of its downstream targets including cytokines like IL-1 $\beta$ , IL-6, and IL-8. These cytokines are upregulated in EOC patients and their increased levels correlate with decreased survival [64–71]. The inflammatory mediators like cytokines, chemokines, growth factors, and prostaglandins secreted by the transformed epithelial cells further promote a pro-inflammatory environment, which can reprogram the surrounding cells to form the TME. The TME is mainly composed of endothelial cells, cancer associated fibroblasts (CAFs), adipocytes, tumor associated macrophages (TAMs), regulatory T-cells, pericytes, infiltrated immune cells such as neutrophils, lymphocytes, and various other cells that further secrete growth factors and cytokines which potentiate tumor progression (Figure 2, Table 1). Furthermore, OC-initiating cells (OCICs) have been identified in tumors and ascites that exhibit stem cell like properties and are capable of forming tumors [65,66,72]. Cytokines can promote self-renewal of CD133<sup>+</sup> OCICs to potentiate tumor progression [73].



**Figure 2.** Inflammatory mediators contributing to EOC progression, metastasis, and angiogenesis. EOC cells produce ROS, chemokines, cytokines, and growth factors that can: (1) Lead to recruitment of immune cells like Dentric cells (DC), Natural killer cells (NK), Tumor associated macrophages (TAMs), and T-regulatory (Treg) cells into the TME, which generate additional cytokines, ROS, and growth factors, resulting in chronic inflammation. (2) Stimulate the tumor cells themselves, the TAMs, and the surrounding fibroblasts (also known as cancer associated fibroblasts or CAFs) to proliferate and secrete growth factors like TGF- $\beta$  and FGF that stimulate production of integrins and Matrix Metalloproteins (MMPs), resulting in migration of the tumor cell via degradation of the extra cellular matrix (ECM). (3) Stimulate endothelial cells (EC) to produce growth factors like PDGF and EGF and factors like VEGF that stimulate angiogenesis. The double arrows indicate that the cells are a source of the factor as well as stimulated by it.

The innate immune response can prevent tumorigenesis by recognizing and eliminating transformed cells. However, chronic inflammation can contribute to the ability of premalignant cells to evade apoptosis, escape the immune surveillance, and continue to grow, resulting in tumor formation. As mentioned, EOC can originate from either distal FT or ovarian epithelial cells. Since both the ovary and fimbria are exposed to the same ME, exposures reviewed here are relevant to initiation in either tissue. [74]. In this section we will review the role of OS and some specific pro-inflammatory mediators and signaling pathways in the initiation and progression of EOC.

### 3.1. ROS and Oxidative Stress

ROS plays an important role in the normal female reproductive cycle, from affecting maturation of the oocyte to ovulation, apoptosis of cells in corpus luteum, and embryo development [75]. Ovulation results in increased levels of DNA damage in the FT epithelium that is likely a result of the ROS generated during ovulation or the ovulation-associated increase in numbers of infiltrating macrophages in the FT [17]. Additionally, during infection and inflammatory responses immune and damaged cells produce ROS resulting in continuous exposure of the ovaries, FTs, and peritoneal cavity to ROS [76–78]. ROS exposure could potentially lead to epithelial cells in the ovary and FT undergoing transformative changes, as has been demonstrated for ovarian surface epithelium cells grown in 3D culture [79]. Elevated ROS and RNS levels beyond the level that cells can neutralize results in OS. Increased OS results in DNA damage, activation of signaling cascades, and epigenetic alterations.

DNA damage in a cell results in stimulation of DNA damage repair pathways. These repair pathways can be inactivated or be erroneous, which results in increased genotoxic stress and mutated DNA. Secretory tubal epithelial cells in the FT, a cell of origin for HGSC, are particularly susceptible to genotoxic injury with persistent DNA damage that could lead to mutation and STIC formation [80].

Mutations in tumor oncogenes and suppressors result in overexpression, constitutive activation of the protein, loss of expression, or expression of nonfunctional proteins, resulting in a transformed cell. Follicular fluid may have transformative properties as it has been demonstrated that bathing fimbriae with follicular fluid containing high levels of ROS results in increased levels of DNA damage. Bathing fimbriae that have loss of p53 and Rb with this follicular fluid results in evasion of apoptosis and cells with persistent DNA damage [81].

ROS can activate pro-survival intracellular tyrosine phosphorylation signaling cascades, mainly regulated by the MAPKs and redox sensitive kinases. Activation of c-Jun, JNK, ERK (extracellular signal-regulated kinase), and p38-MAPK signaling cascades results in upregulation of cell cycle proteins that enhances proliferation. Activation of JNK can also activate NF- $\kappa$ B, which can suppress apoptosis. The MAPK pathway inhibits apoptosis and regulates differentiation. When activated in transformed cells these pathways are important for tumor initiation. ROS affects redox sensitive factors like thioredoxin, which is also found elevated in OC cell lines [82]. Thioredoxin is involved in redox regulation of transcription factors such as NF- $\kappa$ B, NRF2, forkhead box class O (FOXO) proteins, reducing factor-1 (ref-1), and hypoxia inducible factor (HIF-1 $\alpha$ ), thereby increasing their binding to the DNA. Most of these transcription factors promote tumor growth and progression by regulating expression of genes that affect cell survival and growth [83,84]. For example, FOXO, NRF2, and ref-1 transcription factors upregulate transcription of anti-oxidant proteins that scavenge free radicals and allow survival of damaged or transformed cells [85]. HIF-1 $\alpha$  upregulates the antiapoptotic factor, bcl-2 as well as vascular endothelial growth factor (VEGF), a factor important for angiogenesis.

OS has also been shown to facilitate epigenetic mechanisms in many cancers, including EOC [86]. Innate immune-mediated inflammation drives epigenetic silencing of tumor suppressor genes (TSGs) [87]. At sites of inflammation high levels of OS result in oxidative DNA damage that is recognized by the mismatch repair proteins mutS homolog MSH2 and MSH6. MSH2 and MSH6 then recruit epigenetic silencing proteins, including DNA methyltransferase 1 (DNMT1) to the sites of damage [88]. In an in vivo model of inflammation-driven colon tumorigenesis this early recruitment to sites of oxidative DNA damage results in permanent methylation of TSGs in tumors that form at the sites of inflammation [89]. While such a mechanism has not been directly proven in EOC models, Sapoznik et al. have demonstrated that exposure to follicular fluid or inflammation can induce Activation-Induced Cytidine Deaminase (AID) in fallopian tube epithelial cells, which results in epigenetic and genetic changes, increase in DNA damage and genotoxic stress and may be a contributing factor to EOC [90].

### 3.2. TNF- $\alpha$

The cytokine TNF- $\alpha$  plays an important role in the process of ovulation and in removal of damaged corpus luteum. TNF- $\alpha$  ligand and its receptors, TNFRI and TNFRII are upregulated in ovarian tumors compared to normal ovarian tissue and high levels of TNF- $\alpha$  are found in ascites from OC patients [91–93]. OC cells have also been shown to secrete high levels of TNF- $\alpha$  as compared to normal ovarian epithelial cells resulting in autocrine upregulation of TNF- $\alpha$  mRNA and in expression of other pro-inflammatory cytokines, chemokines, and angiogenic factors like IL-6, M-CSF, CXCL2, CCL2, and VEGF [93,94]. Kellie et al. have shown using mouse models that TNF- $\alpha$  stimulates IL-17 production via TNFRI resulting in myeloid cell recruitment to the ovarian TME and increased tumor growth [95]. TNF- $\alpha$ , also upregulates AID transcript levels which can contribute to genotoxic stress [90].

### 3.3. IL-6

The cytokine IL-6 has been associated with poor survival in OC and is emerging as a potential therapeutic target for EOC [67,68,96,97]. IL-6 is normally produced by ovarian epithelial and OC cells. Macrophage migration inhibitory factor (MIF), EGF, and Transglutaminase secreted by OC cells can stimulate IL-6 production via activation of NF- $\kappa$ B [98–100]. IL-6 increases proliferation of OC cells by



facilitating their exit from G1 into S phase of the cell cycle and by activation of the MAPK-ERK-Akt (protein kinase B) growth promoting signaling pathway [101]. ERK activation can promote formation of ascites by increasing the migration of tumor cells [70]. IL-6 production by M2 macrophages present in ascites in later stages of EOC can also stimulate cancer cell proliferation via STAT3 activation [102]. High levels of IL-6 can result in immune suppression by downregulation of IL-2, which stimulates Teff cell production [103]. IL-6 also stimulates production of Metallomatrix proteins (MMPs) in OC cells, which increases their invasive properties and promotes tumorigenesis [101,104].

### 3.4. IL-8

IL-8 a member of C-X-C chemokine family is present in the preovulatory follicle [105] where it may play a role in increasing leukocyte infiltration [106]. It is also elevated in ovarian cysts and in OC patients compared to healthy controls [107,108]. IL-8 has been found to be present in significantly higher levels in the ascites of patients with OC in comparison to patients with benign gynecological disorders [109]. Increased IL-8 expression has been associated with poor prognosis in OC patients [107]. Treatment of EOC cells with IL-8 results in their increased proliferation, which is accompanied by an increase in cyclins B1 and D1 and is dependent on phosphorylation of Akt and ERK [110]. Cyclins B1 and D1 are important for cell cycle progression, and an increase in their expression leads to increased cell growth. On the other hand, two independent studies have demonstrated that IL-8 inhibits EOC growth by increasing neutrophil infiltration [111,112].

### 3.5. Lyophosphotidic Acid (LPA)

LPA is a phospholipid that binds to and activates the endothelial differentiation gene (Edg) family of receptors. LPA is present in ovarian follicular fluid and it stimulates IL-6 and IL-8 production in the corpus luteum [113,114]. OC cells have been shown to produce LPA, which functions like a growth factor [115–119]. Plasma and ascites of OC patients have elevated levels of LPA that contribute to OC progression via upregulation of COX-2 and MMP2 [115,120,121]. LPA can bind to LPA<sub>2</sub> receptor and induce expression of IL-6 and IL-8 via activation of NF- $\kappa$ B and AP-1 in OC cell lines [122]. It can induce ROS dependent Akt and ERK phosphorylation and inhibition of LPA can increase apoptosis of EOC cells [123]. ERK phosphorylation can induce phosphorylation of HIF-1 $\alpha$ , which then can upregulate VEGF and promote tumorigenesis. Another group demonstrated that stimulating EOC cells with ether-linked LPA resulted in their increased proliferation and survival by increased synthesis of DNA and activation of Akt via PI3K, which contributes to tumor progression [124].

### 3.6. Prostaglandins and COX-1 and COX-2

Prostaglandins are secreted in the ovary, FT, and uterus. They are important for maturation of the oocyte and facilitate the movement of the FT so that the mature oocyte can move from the ovary to the uterus. In the uterus prostaglandins help regulate and maintain uterine blood flow. COX-1 and COX-2 are enzymes that catalyze the production of prostaglandins from arachidonic acid and are overexpressed in OC patients [22,125,126]. High COX levels positively correlate with increased cell proliferation, angiogenesis, and malignancy in ovarian tumors [126,127]. COX-1 and COX-2 are normally involved in the acute inflammatory response but can become dysregulated in chronic inflammatory or TMEs. Obermajer et al. have demonstrated that prostaglandins produced by COX-2 can stimulate production of CXCR4 and its ligand Stromal cell derived factor 1 (SDF1) CXCL12 in myeloid derived suppressor cells (MDSC), which stimulates them to migrate towards OC ascites [128]. MDSCs inhibit the proliferation and differentiation of T cells, resulting in overall immune suppression, which allows the tumor cells to escape immune surveillance and continue to grow. Genetically engineered mouse models of EOC; one harboring the *p53* and *Rb* deletion and other the *KRAS*<sup>G12D</sup> mutation and *Pten* deletion, demonstrate increased COX-1 levels, thus suggestive that COX-1 could be used as a potential biomarker and therapeutic target for EOC [129]. Further when

COX-1 was inhibited in EOC cells, it led to reduction in prostacyclin (a type of prostaglandin) synthesis and reduced tumor growth by enhanced apoptosis [130].

#### 4. Inflammation and EOC Angiogenesis

Angiogenesis is required for the growth of both primary and metastatic tumors [131]. The process of angiogenesis is a complex multi-step process reviewed previously [132]. It is regulated by a balance between pro-angiogenic and antiangiogenic factors. Hypoxic and ischemic areas are present at sites of inflammation and also in tumors mainly due to obstruction of local blood vessels, differences in pace of growth of blood vessels and growth of the tumor and/or infiltration of immune cells. Macrophages accumulate at hypoxic sites and alter their gene expression profiles in response to the hypoxic conditions. One of the important genes for angiogenesis that is upregulated by hypoxia is VEGF [133,134]. The rate-limiting step in angiogenesis is VEGF signaling in endothelial cells (ECs) [135]. VEGF functions via tyrosine kinase receptors VEGF-1 and VEGF-2 and promotes migration, survival, proliferation of ECs, and formation of new blood vessels [136–138]. Many of the inflammatory mediators discussed so far are also involved in promoting angiogenesis in EOC as detailed below (Figure 2, Table 1).

##### 4.1. TNF- $\alpha$

TNF- $\alpha$  creates a pro-inflammatory TME and has also been associated with promoting angiogenesis. It has been hypothesized that TNF- $\alpha$  induces the production of soluble factors that promote tumor angiogenesis. Culture supernatants from TNF- $\alpha$  expressing cells induce the growth of mouse lung endothelial cells in vitro while culture supernatants from TNF- $\alpha$  lacking cells do not exert the same effect [94]. In pituitary adenomas TNF- $\alpha$  is known to induce VEGF that in turn induces CXCL12 [139,140]. VEGF and CXCL12 synergistically induce angiogenesis in EOC [141]. Mice injected with OC cells lacking TNF- $\alpha$  have reduced vascular density in their tumors and reduced formation of blood vessels in the peritoneal deposits. These mice also did not have accumulation of ascetic fluid suggesting the importance of TNF- $\alpha$  in angiogenesis and EOC progression [94].

##### 4.2. IL-6

In physiological conditions, IL-6 is involved in angiogenesis in the ovary during the development of ovarian follicles [142]. IL-6 induces the phosphorylation of STAT3 and MAPK in ovarian endothelial cells thereby enhancing their migratory ability, a key step in angiogenesis [143]. As explained before, OC cells also secrete increased amounts of IL-6. Some OC cells also secrete an alternative splice variant of IL-6R $\alpha$ , the soluble form sIL-6R, which consists of only the ectodomain of the transmembrane receptor. By a process called trans-signaling, the sIL-6R-IL-6 complex initiates signaling in cells in the ME that do not express the transmembrane receptor facilitating angiogenesis [144].

##### 4.3. IL-8

Several studies have clearly established the role of IL-8 in promoting angiogenesis. Hu et al., demonstrated that IL-8 plays a role in angiogenesis using a rat sponge model [145]. IL-8 was also able to induce angiogenesis in the rat cornea, which is normally avascular [146]. As explained in the previous section, there are several sources of IL-8 in ovarian TME. Overexpression of IL-8 in A2780 (non-IL-8 expressing) OC cells has been shown to increase the expression of VEGF, MMP-2, and MMP-9; while depletion of IL-8 in SKOV3 (IL-8 expressing) cells has been shown to reduce VEGF, MMP-2, and MMP-9 [110]. The process of angiogenesis involves degradation of extracellular matrix components and proliferation and migration of endothelial cells. MMPs are a family of endopeptidases that breakdown components of extracellular matrix and have been implicated in angiogenesis [147]. Because of the importance of VEGF and MMPs in angiogenesis these findings suggest that IL-8 in the ovarian TME will promote the formation of new blood vessels in EOC. Targeting IL-8 using mouse models reduces EOC growth and decreases angiogenesis [112].

**Table 1.** Role of inflammatory mediators in different stages of tumor progression.

Inflammatory Mediators	Secreting Cell Type	Stages in Tumor Progression			
		Initiation and Progression	Angiogenesis	Metastasis	Chemoresistance
TNF- $\alpha$ ligands, TNFRI, TNFRII	OC cells, infiltrating monocytes, macrophages	$\uparrow$ autocrine production of TNF- $\alpha$ and IL-6, M-CSF, CXCL2, CCL2 [93,94] and AIDS mRNA level [90]	$\uparrow$ VEGF, VEGF $\uparrow$ CXCL12 and promotes angiogenesis [139–141]	$\uparrow$ TGF- $\alpha$ secretion by stromal fibroblasts which promote peritoneal metastasis [148] Enhances migration of OC cells towards CXCL12 [149,150]	
IL-6	Ovarian epithelial cells, OC cells, M2 macrophages, mesothelial cells, TAMS, ascites	$\uparrow$ Proliferation by promoting G1 to S transition and MAPK-ERK-Akt activation and STAT3 activation [101,102] $\downarrow$ IL-2, resulting in immune suppression [103]	Induces STAT3 and MAPK phosphorylation which enhances migration of endothelial cells [143] sIL-6R-IL-6 facilitates angiogenesis in cells lacking IL-6 receptor [144]	Stimulates production of MMPs in OCs which $\uparrow$ invasion and migration [101,104] $\uparrow$ IL-6 in ascites enhances invasion via JAK-STAT signaling [151]	$\downarrow$ Caspase-3 cleavage and makes OC cells resistant to cisplatin and paclitaxel [152] $\uparrow$ Expression of MDR1, GSTpi, Bcl-2, Bcl-xL, and XIAP [152]
IL-8	Pre-ovulatory follicles, OC cells, ascites	$\uparrow$ Proliferation by $\uparrow$ cyclin B1 and cyclin D1 via pAkt [110]	$\uparrow$ Expression of VEGF, MMP-2, MMP-9 promoting angiogenesis [110]	Activates TAK1/ NF- $\kappa$ B via CXCR2 [153]	Blocks TRAIL induced apoptosis to promote resistance [154]
LPA	Follicular fluid, corpus luteum, OC cells, ascites	$\uparrow$ IL-6 and IL-8 via NF- $\kappa$ B and AP-1 [113,114,122] $\uparrow$ COX-2 AND MMP2 [115,120,121] $\uparrow$ phosphorylation of Akt and ERK resulting in increased cell cycle [123,124]	$\uparrow$ Expression of VEGF via Myc and Sp-1 [155]	$\uparrow$ urokinase, which results in degradation of basemembrane protein to promote metastasis [156,157]	
Prostaglandins, COX-1 and COX-2	Ovary, FT, uterus, MDSCs	$\uparrow$ CXCR4 and SDF1 in MDSCs resulting in immune suppression [128]	$\uparrow$ Bcl-2 and blood vessel formation [158,159]		$\uparrow$ Bcl-2, thus inhibiting apoptosis in lung, colon, breast and prostate cancers [158,159]
TGF- $\beta$ and EGF	OC cells, CAFs			TGF- $\beta$ $\uparrow$ VCAN, which activates NF- $\kappa$ B and $\uparrow$ MM-9 [160]	$\uparrow$ EGF protects cells from cisplatin-induced apoptosis [161]. Inhibiting TGF- $\beta$ sensitizes resistant cells [162]

#### 4.4. LPA

In addition to playing a role in initiation, and progression, LPA has also been implicated in angiogenesis in OC. LPA has been shown to induce transcriptional activation of VEGF in EOC cell lines [163]. Transcriptional activation of VEGF primarily occurs through HIF-1 $\alpha$  under oxygen limiting conditions in Hep3B hepatocellular carcinoma cells [164]. LPA mediated induction of VEGF expression has been shown to be independent of HIF-1 $\alpha$  in EOC cell lines. Transition metal cobalt treatment also leads to stabilization of HIF1 $\alpha$  similar to hypoxia. Combination treatment of EOC cells with cobalt and LPA additively increased VEGF production suggesting the effect of two different pathways [155]. LPA activates c-Myc and Sp-1, which induce VEGF expression through consensus binding sites in the VEGF promoter that have been implicated in HIF $\alpha$  independent induction of VEGF [155].

### 5. Inflammation and EOC Metastasis

Tumor metastasis is the major cause of mortality in most cancers, including EOC. Most EOC patients are diagnosed at an advanced stage when the cancer has already metastasized [165]. Dissemination of cancer cells to distant sites is a complex multi-step process called the invasion-metastasis cascade and is reviewed in detail in previous papers [166–168]. Briefly, some major steps in metastasis are—invasion through the basement membrane, intravasation into the lymphatics and circulation, survival of disseminating cancer cells in circulation, extravasation into surrounding tissues, colonization, and finally, formation of micro and macro metastases. However, unlike other epithelial malignancies, EOC has a different pattern of metastasis. EOC cells directly shed from the primary tumor into the peritoneal space and disseminate to organs in the peritoneal cavity. One of the prerequisites for cancer cells to metastasize is to undergo a process called epithelial to mesenchymal transition (EMT) where they lose their ability to attach to the basement membrane and acquire a mesenchymal phenotype and characteristics. Several recent evidences have indicated that the TME aids tumor cells to acquire these properties facilitating the metastatic cascade. An example of the ME promoting metastasis is the presence of STICs in the distal part of the FT, which shares its ME with ovary. Yang-Hartwich et al. have demonstrated that granulosa cells in the ovary secrete SDF-1 (stromal cell-derived factor 1) [169]. SDF-1 functions as a chemoattractant and recruits malignant FT cells to the ovary suggesting that the ovary is a primary site of metastasis, not the primary tumor site. Russo et al. demonstrated that loss of PTEN (phosphatase and tensin homolog) by the malignant FT cells and upregulation of WNT4 (wingless-related MMTV integration site 4) is crucial for initial metastasis to the ovary thereby supporting the tubal origin of EOC and the ovary as the primary site of metastasis [170]. The cells that make up the TME also secrete various inflammatory mediators, which facilitate progression and metastasis of OC cells (Figure 2, Table 1). These factors enable tumor metastasis by deregulating signal transduction pathways. Examples include the PI3-Akt and RAS-ERK pathways, which control migration and invasion through downstream effectors like Rho family GTPases, extracellular proteases, integrins, matrix associated proteins like focal adhesion kinases (FAK), and transcription factors like ETS2 and AP-1 [171–173]. Robinson-Smith et al. demonstrated that peritoneal inflammation correlated with dissemination of cancer cells from the ovaries in SCID mice. Augmenting the inflammatory response using thioglycolate accelerated ascites formation and metastasis while suppressing the inflammation using acetyl salicylic acid impeded ascites formation and reduced metastasis. This inflammation-induced metastasis of OC cells was found to be primarily mediated by macrophages and not neutrophils or NK cells [174]. As explained in one of the previous sections a pro-inflammatory environment can be created in the peritoneum due to secretion of cytokines like IL-6 and TNF- $\alpha$  by adipose cells [31]. Omentum, the primary site of metastasis of OC, is largely composed of adipose cells. In addition to adipocytes, omentum also consists of blood and lymph vessels, immune cells, and stromal cells [175]. Adipocytes have been shown to increase migration, invasion, and proliferation of EOC cells. Upregulation of SUSD2 a secreted tumor suppressor by adipocytes by guadecitabine treatment reduced EOC migration and invasion. This finding suggests that epigenetic changes in the stromal cells in addition to EOC cells can facilitate EOC

metastasis [176]. Omentum has aggregates of immune cells around the vasculature commonly referred to as milky spots [177]. Melanoma, lung carcinoma, ovarian carcinoma, and mammary carcinoma cell lines have been shown to specifically metastasize to the immune cell aggregates in the omentum when injected intraperitoneally into C57BL/6 mice [178]. These milky spots in the omentum have also been shown to facilitate metastatic colonization of the OC cells. Clark et al. have suggested that both adipocytes and milky spots have specific and important roles in metastatic colonization of OC cells [179]. These evidences imply that omentum potentially provides a good niche for the growth of ovarian cancer cells. Here we will specifically discuss how inflammatory mediators promote tumor metastasis in EOC.

### 5.1. ROS

EOC cells produce a large amount of ROS [180]. Loss of E-cadherin is one of the characteristic features of tumor cells with increased ability to migrate and invade. Wang et al. demonstrated that ROS leads to HIF $\alpha$  mediated activation of lysyl oxidase. Lysyl oxidase was shown to inversely correlate with E-cadherin expression promoting migration and invasion in EOC cells [181]. Tumor cells treated with sub-lethal doses of H<sub>2</sub>O<sub>2</sub> failed to attach to the extracellular matrix components fibronectin and laminin and had increased metastatic colonization of lung, thereby establishing a role for ROS in tumor cell metastasis [182].

### 5.2. TNF- $\alpha$

TNF- $\alpha$  provides a good example of how interactions between cancer and stroma aid in OC metastasis. Ascitic fluid and OCs contain a large number infiltrating macrophages in part because OCs constitutively produce M-CSF, which functions as a chemoattractant for monocytes [183]. These infiltrating monocytes produce many cytokines one of which is TNF- $\alpha$  [184,185]. OC cells also have elevated TNF- $\alpha$  expression that is regulated by DNA hypomethylation and chromatin remodeling of the TNF- $\alpha$  promoter. Increased TNF- $\alpha$  produced by OC cells and macrophages stimulates increased expression of TGF- $\alpha$  in stromal fibroblasts. TGF- $\alpha$  secreting stromal fibroblasts promote peritoneal metastasis of OC via EGF receptor signaling [148].

Furthermore, in EOC cells and clinical biopsies TNF- $\alpha$  expression correlates with one of the most commonly expressed cytokine receptors CXCR4. TNF- $\alpha$  stimulation of EOC cells enhanced their migration toward the only CXCR4 ligand, CXCL12. Stimulation of EOC cells by CXCL12 induced mRNA and protein expression of TNF- $\alpha$ . Therefore, a positive feedback loop has been suggested where in CXCL12 induced TNF- $\alpha$  potentially acts on the cancer cells and induces CXCR4 expression thereby enhancing tumor cell migration [149,150].

### 5.3. IL-6

IL-6 has also been implicated in metastasis of OC. Elevated levels of IL-6 found in serum and peritoneal fluid of EOC and OC patients have many sources [186–188]. Mesothelial cells in the peritoneum, TAMs, and EOC cells all secrete IL-6 [67]. M2 polarized macrophages in the ovarian TME induce proliferation and invasion of EOC cells by secretion of IL-6 [189]. Increased IL-6 present in ascites from OC patients enhanced the invasive ability of OC cells via the JAK-STAT signaling pathway. Canonically IL-6 signaling occurs by binding of the ligand to its transmembrane receptor IL-6R $\alpha$ . The effect of IL-6 on invasion of OC cells correlated with their IL-6R expression [151]. Because through trans-signaling, the sIL-6R–IL-6 complex initiates signaling in cells that do not express the transmembrane receptor [144], we hypothesize that IL-6 produced by macrophages could also promote invasion of OC cells similar to the mechanism of induction of angiogenesis.

### 5.4. IL-8

Increased proliferation, anchorage independent growth, and angiogenic potential are some prerequisites for cells to metastasize. IL-8 increases the proliferation of OC cells and upregulates VEGF



and MMP2 and 9 via activation of NF- $\kappa$ B, which results in enhanced invasive phenotype of OC cells. IL-8 has been shown to activate TAK1/NF- $\kappa$ B signaling via CXCR2, thereby facilitating the seeding and growth of OC cells in the peritoneal cavity during metastasis [153].

### 5.5. LPA

LPA promotes proliferation, survival, and metastasis of EOC cells by inducing the expression of c-Myc, VEGF, IL-8, MMPs and COX-2 [163,190–193]. LPA acts through its receptors LPAR1-3, which are members of G-protein coupled receptor superfamily. Invasive EOC cells have significantly higher expression of LPAR1 in comparison to non-invasive cell lines and LPA induces EOC cell invasion specifically through LPAR1 and not through LPAR2 or LPAR3 [194]. It can also induce secretion of urokinase in EOC cells, which has been shown to play a role in metastasis and its high levels correlate with advanced OC and poor survival in patients. LPA has been shown to increase promoter activity, mRNA levels, protein levels, and enzyme activity of Urokinase plasminogen activator (uPA) possibly via the edg-4 LPA receptor [156]. uPA is involved in converting plasminogen to plasmin, which facilitates the degradation of basement membrane and extracellular membrane proteins like fibronectin aiding in metastasis [157].

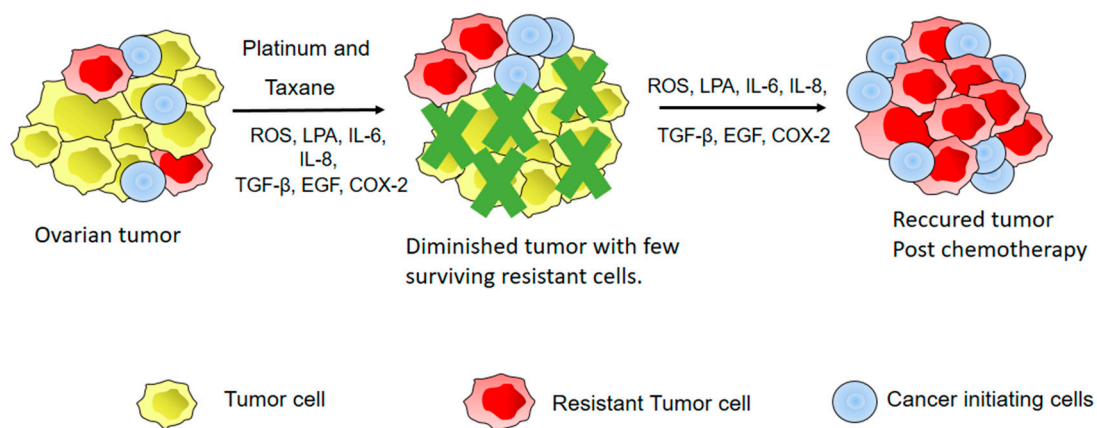
### 5.6. TGF- $\beta$

TGF- $\beta$  initiates signaling by dimerization of serine/threonine kinase receptors. The dimerization of receptors results in their phosphorylation, which then relays signals downstream via SMAD dependent and SMAD independent pathways. Phosphorylation by the TGF- $\beta$  receptor causes R-SMADs to bind to Co-SMAD and translocate to the nucleus, where they activate transcription of genes that promote invasion, migration. Bone morphogenic proteins (BMPs) are cytokines that belong to TGF- $\beta$  family and have been associated with progression of many different cancer types. Their mechanism of promoting tumor progression depends on the TME in which the cancer grows and their mode of metastatic spread [195]. Specifically, BMP-2 overexpression has been associated with poor prognosis in OC [196]. Additionally, TGF- $\beta$  could potentially modify the TME to promote tumorigenesis. Veriscan (VCAN), an extracellular matrix associated protein, was upregulated by TGF- $\beta$  through TGF- $\beta$  receptor II (TGFB2) and SMAD signaling making the EOC cells more aggressive. Increased VCAN expression enhanced motility and invasion of EOC cells by activating NF- $\kappa$ B signaling, increased expression of MMP-9, and hyaluronidase mediated motility receptor [160]. CAFs have higher expression of TGF- $\beta$  receptors in comparison to normal ovarian fibroblasts and EOC cells suggesting that CAFs within the TME are more responsive to TGF- $\beta$  than the other cell types [160].

## 6. Inflammation and EOC Chemoresistance

The standard treatment for EOC patients is cytoreductive surgery followed by platinum/taxane-based chemotherapy [197]. The main obstacle in treatment of EOC patients is development of chemoresistance. Resistance to chemotherapy can be either intrinsic or acquired. Inherent gene expression patterns harbored by chemo-naïve tumor cells contribute to intrinsic resistance. Acquired resistance is a consequence of different alterations induced after exposure to chemotherapeutic agents [198]. Different mechanisms, including increased drug efflux, decreased uptake of the drug, inactivation of the drug, increased DNA repair, and reduced apoptotic response, have been implicated in development of platinum resistance [199]. Several recent studies have demonstrated that the TME contributes to both intrinsic and acquired resistance. One type of intrinsic drug resistance influenced by the TME is referred to as environment mediated drug resistance (EMDR). In EMDR, factors and cells present in the TME activate diverse signaling events, transiently protecting the tumor cells from undergoing apoptosis in response to chemotherapeutic agents [200,201]. Another type of drug resistance induced by cytokines, chemokines, and growth factors secreted by fibroblast cells in the tumor stroma is called soluble factor mediated drug resistance (SFM-DR). A good example of SFM-DR is IL-6 mediated drug resistance in multiple myeloma. IL-6 is important for growth of multiple

myeloma cells. IL-6 activates STAT3 signaling in these cells and protects them from Fas mediated apoptosis by upregulating antiapoptotic protein Bcl-X<sub>L</sub> [202]. Myeloma cells that produced IL-6 in an autocrine manner were found to be resistant to dexamethasone induced apoptosis while non-IL-6 producing cells were sensitive [203]. Cell adhesion mediated drug resistance (CAM-DR) occurs due to adhesion of tumor cells to extracellular matrix components like laminin, collagen, and fibronectin or due to fibroblasts present in the tumor stroma [204]. An example of this type of resistance is when drug sensitive myeloma cells were adhered to an extracellular matrix component fibronectin, they exhibited a reversible drug resistant phenotype which was not due reduced drug accumulation or increase in antiapoptotic proteins like Bcl-X<sub>L</sub> [201]. Here we will discuss specific inflammatory mediators and their role in OC chemoresistance (Figure 3).



**Figure 3.** Inflammatory mediators contribute to chemoresistance of EOC. A combination of platinum and taxane drugs is currently used as chemotherapy for OC. ROS, Lyophosphotidic Acid (LPA), cytokines, and growth factors like TGF- $\beta$  and EGF increase tumor cell survival by upregulating antiapoptotic genes, by stimulating stemness and proliferation of cancer initiating cells, by increasing repair of damaged DNA, or by increasing efflux of the drug. The resistant tumor cells and the cancer initiating cells can then proliferate under the influence of growth factors and cytokines resulting in a recurrent chemoresistant tumor.

### 6.1. ROS

ROS are abundant in the pro-inflammatory TME. Malignant EOC tissues have been shown to have 96% higher ROS levels than normal controls [205]. OC stem like cells or OCICs are more drug resistant and responsible for relapse of chemoresistant tumors [66]. OCICs produce ROS and superoxide. This ROS induces the expression of peroxisome proliferator-activated receptor-gamma coactivator (PCG)-1 $\alpha$ , which regulates mitochondrial biogenesis and is required for expression of detoxifying enzymes [206,207]. PCG1 $\alpha$  increases the aldehyde dehydrogenase (ALDH) activity and expression of multidrug resistance gene (MDR1). MDR1 is an ATP dependent transporter that has been associated with efflux of platinum based drugs from OC cells contributing to platinum resistance. Scavenging ROS reduced expression of PCG1 $\alpha$  and drug resistant related genes thereby linking ROS to development of chemoresistance [207].

### 6.2. IL-6

IL-6 in the OC TME is associated with increased chemoresistance. Wang et al. demonstrated that autocrine production of IL-6 by EOC cells makes them resistant to cisplatin and paclitaxel by causing decreased proteolytic cleavage of capase-3. Paclitaxel resistant EOC cells have increased expression of IL-6 and one of its downstream effectors STAT3 [208,209]. IL-6 producing OC cells also had increased expression of multidrug resistant genes MDR1 and GSTpi and anti-apoptotic genes

Bcl-2, Bcl-xL, and XIAP, suggesting that IL6 promotes drug resistance by increasing drug efflux and reducing apoptosis [152].

### 6.3. IL-8

IL-8 blocks TRAIL-induced apoptosis and reduces caspase cleavage in EOC cell lines by decreasing the expression of death receptor (DR) 4 [210]. TRAIL is a cell death inducing ligand that belongs to the TNF superfamily and has been shown to induce apoptosis specifically in tumor cells and not in nontransformed cells [211,212]. Combination of TRAIL and the chemotherapeutic drugs—cisplatin, doxorubicin, and paclitaxel has been shown to induce apoptosis in chemoresistant EOC cell lines by causing increased caspase and PARP cleavage [154]. This finding suggests that IL8 may contribute to chemoresistance by blocking TRAIL.

### 6.4. LPA

LPA has been shown to contribute to platinum resistance by preventing cells from undergoing cisplatin-induced apoptosis without affecting their proliferation rate. The mechanism of how LPA inhibits apoptosis in EOC cells in response to cisplatin is not yet clearly understood [161].

### 6.5. TGF- $\beta$ and EGF

Recurrent OC show significantly higher expression of TGF- $\beta$ 1 and TGF- $\beta$ 3 in comparison to primary tumors and normal ovary tissue [213]. Inhibition of TGF- $\beta$  by the inhibitor LY2109761 sensitizes resistant SKOV3 cells to cisplatin suggesting that TGF- $\beta$  contributes to the development of platinum resistance in EOC cells [162]. Cisplatin resistant A2780P cells had hypomethylation and upregulation of TGFBR2 confirming the involvement of the pathway in acquisition of platinum resistance [214]. An elevated level of EGF receptor (EGFR) has also been associated with poor prognosis in OC patients [215]. EGF has been shown to stimulate the growth of EOC cells expressing EGFR and alters their cell cycle distribution [216]. EGF similar to LPA has been shown to protect EOC cells from undergoing cisplatin induced apoptosis [161].

### 6.6. COX-2

In addition to being associated with tumor initiation and progression, COX-2 has also been associated with chemoresistance. Ferrandina et al. reported that a statistically significant higher percentage of primary OC patients unresponsive to platinum-containing chemotherapy were positive for COX-2 than responsive patients (84.6% versus 34.6%, respectively) [217]. The percentage of positive COX-2 staining per tumor area in COX-2 positive patients ranged from 15 to 45%. The results from this study suggest that COX-2 levels may influence the response of patients to different chemotherapy regimens, but the sample size of this study was small and the results need to be confirmed in a larger group of patients. Furthermore, this association needs to be corroborated biochemically [217]. In both patients groups undergoing cytoreductive surgery and explorative laparotomy, COX-2 expression was higher in nonresponders [218]. Using lung, colon, and prostate cancer models, COX-2 has been shown to induce Bcl-2 and promote tumor growth by facilitating the formation of new blood vessels [158,159]. These findings suggest that COX-2 may contribute to chemoresistance by inhibiting apoptosis and promoting angiogenesis in OC as well.

## 7. Treatment Strategies Targeting Inflammatory Mediators in EOC

As discussed, development of resistance to available chemotherapeutic drugs remains the major obstacle in management of OC patients. While several immunotherapies have been developed to improve the antitumor response of T-cells and/or modulate the immune response, here we will discuss EOC treatment strategies that specifically target the inflammatory mediators that have been reviewed above.

A monoclonal antibody directed at VEGF, bevacizumab, has been widely studied and is a promising target in EOC [219]. Bevacizumab is a recombinant humanized monoclonal antibody and has been approved by the FDA for treatment of metastatic breast, non-small cell lung, and colorectal cancer. Phase II clinical studies have shown that it is active in treatment of recurrent OC patients [220]. OCEANS trial was a randomized phase III clinical trial that evaluated the safety and efficacy of bevacizumab in combination with gemcitabine and carboplatin (GC) in comparison with GC alone in recurrent platinum sensitive ovarian, primary peritoneal, or FT cancer. This trial demonstrated that bevacizumab was able to prolong the PFS in platinum-sensitive recurrent EOC patients [221]. In addition to OCEANS, GOG218, and ICON7 have also shown that bevacizumab prolongs the PFS in OC patients confirming the promise this therapeutic target holds for management of OC [222,223].

We have discussed some mechanisms by which the pro-inflammatory cytokine TNF- $\alpha$  promotes OC metastasis and angiogenesis making it a good target for development of therapeutic agents. The safety profile and biological activity of a monoclonal anti-TNF- $\alpha$  antibody, Infliximab was assessed in a clinical study consisting of patients with advanced solid tumors, including OC. Infliximab did not have any toxic effects and was well tolerated by these patients. Reduced plasma levels of IL-6 and CCL12 in these patients was observed 24 h and 48 h after administration of Infliximab, while neutralization of TNF- $\alpha$  was detected after an hour indicating some biological activity [224]. This response warrants further study of Infliximab as a therapeutic agent for treatment of OC.

IL-6/STAT3 signaling has been implicated at different stages of OC progression and is a promising target although most agents are still in preclinical or early clinical trial stages. Siltuximab, an anti-IL-6 antibody, suppresses IL-6-induced STAT3 phosphorylation and nuclear translocation in OC cell lines. Siltuximab treatment also reduced the level of pro-survival proteins like Bcl-X<sub>L</sub> and Survivin, which are downstream of STAT3. Siltuximab was able to sensitize paclitaxel resistant OC cell lines, but did not show the same effect in vivo [225]. sc144 is a novel small molecule inhibitor has shown significant promise in preclinical studies. sc144 binds gp130, which is a signal transducer in STAT3 signaling. It causes phosphorylation of gp130 leading to its deglycosylation. This abrogates downstream STAT3 phosphorylation and nuclear translocation inhibiting transcription of downstream genes. sc144 has increased potency in EOC cells in comparison to normal epithelial cells and slows down the growth of tumors in xenograft models of EOC [226]. A phase I clinical trial combining carboplatin, the monoclonal antibody Tocilizumab, which blocks IL-6R, and immune enhancer INF- $\alpha$  showed good promise. The EOC patients who received the highest dose of Tocilizumab had increased serum levels of IL-6 and sIL-6R and also showed longer median overall survival [227].

We have discussed the role of TGF- $\beta$  in EOC tumor progression substantiating it as a good therapeutic target. A preclinical study of LY2109761 (TGF $\beta$ RI and TGF $\beta$ RII kinase inhibitor) in combination with cisplatin was conducted by Gao et al. This inhibitor significantly increased apoptosis in cisplatin resistant cells. Combining LY2109761 with cisplatin had antiproliferative effects and increased the rate of apoptosis in parental and cisplatin resistant xenograft models [162]. In triple negative breast cancer, LY2157299 a TGF- $\beta$ 1 receptor kinase inhibitor, prevented recurrence of tumors in xenograft models after treatment with paclitaxel [228]. Early phase clinical trials of LY2157299 in patients with advanced or metastasized pancreatic cancer have been completed. Early phase trials in triple negative metastatic breast cancer, unresectable hepatocellular carcinoma, and metastatic castration resistant prostate cancer are underway [229].

EGF has also been associated with chemoresistance in EOC. Cetuximab, a chimerized monoclonal antibody that targets EGFR, was tested in combination with carboplatin in patients with recurrent platinum sensitive OC. Cetuximab showed modest activity in these patients [230]. Panitumumab, a human monoclonal antibody specific to EGFR, in combination with carboplatin did not improve efficacy or progression free survival in platinum sensitive EOC patients [231].

## 8. Conclusions and Future Perspectives

Several studies in the last decade have associated increased inflammation and inflammatory mediators with increased EOC risk and reduced survival in EOC patients. We have presented published evidence suggesting that inflammation and inflammatory mediators promote ovarian tumorigenesis. However the mechanisms by which the process of inflammation culminates in ovarian tumor initiation need to be further understood. Such links have been established in colon and pancreatic cancer. Understanding these mechanisms is important for developing ways to target inflammatory mediators and reduce OC risk. Furthermore, epidemiological studies of NSAIDs and early clinical trials targeting IL-6 and TNF- $\alpha$  have shown significant promise, thus suggesting that targeting inflammatory mediators as treatment for OC warrants future research.

**Author Contributions:** S.S.S. and S.S. wrote the manuscript. S.S.S., S.S. and H.M.O. edited the manuscript.

**Funding:** This research was funded by National Institute of Environmental Health Sciences grant number R01ES023183 and Ovarian Cancer Research Fund grant number 458788.

**Acknowledgments:** We acknowledge the Leo and Anne Albert Charitable Trust for its ongoing support of ovarian cancer research.

**Conflicts of Interest:** The authors declare no conflicts of interest.

## References

1. Maiuri, A.R.; O'Hagan, H.M. Interplay Between Inflammation and Epigenetic Changes in Cancer. *Prog. Mol. Biol. Transl. Sci.* **2016**, *144*, 69–117. [[PubMed](#)]
2. Siegel, R.L.; Miller, K.D.; Jemal, A. Cancer statistics, 2018. *CA Cancer J. Clin.* **2018**, *68*, 7–30. [[CrossRef](#)] [[PubMed](#)]
3. Grivennikov, S.I.; Greten, F.R.; Karin, M. Immunity, inflammation, and cancer. *Cell* **2010**, *140*, 883–899. [[CrossRef](#)] [[PubMed](#)]
4. Clendenen, T.V.; Lundin, E.; Zeleniuch-Jacquotte, A.; Koenig, K.L.; Berrino, F.; Lukanova, A.; Lokshin, A.E.; Idahl, A.; Ohlson, N.; Hallmans, G.; et al. Circulating inflammation markers and risk of epithelial ovarian cancer. *Cancer Epidemiol. Biomark. Prev.* **2011**, *20*, 799–810. [[CrossRef](#)] [[PubMed](#)]
5. Chou, C.H.; Wei, L.H.; Kuo, M.L.; Huang, Y.J.; Lai, K.P.; Chen, C.A.; Hsieh, C.Y. Up-regulation of interleukin-6 in human ovarian cancer cell via a Gi/PI3K-Akt/NF- $\kappa$ B pathway by lysophosphatidic acid, an ovarian cancer-activating factor. *Carcinogenesis* **2005**, *26*, 45–52. [[CrossRef](#)] [[PubMed](#)]
6. Dhillon, A.S.; Hagan, S.; Rath, O.; Kolch, W. MAP kinase signalling pathways in cancer. *Oncogene* **2007**, *26*, 3279–3290. [[CrossRef](#)] [[PubMed](#)]
7. Schulze-Osthoff, K.; Ferrari, D.; Riehemann, K.; Wesselborg, S. Regulation of NF- $\kappa$ B activation by MAP kinase cascades. *Immunobiology* **1997**, *198*, 35–49. [[CrossRef](#)]
8. Coussens, L.M.; Werb, Z. Inflammation and cancer. *Nature* **2002**, *420*, 860–867. [[CrossRef](#)] [[PubMed](#)]
9. Gajewski, T.F.; Schreiber, H.; Fu, Y.X. Innate and adaptive immune cells in the tumor microenvironment. *Nat. Immunol.* **2013**, *14*, 1014–1022. [[CrossRef](#)] [[PubMed](#)]
10. Wilson, C.B.; Rowell, E.; Sekimata, M. Epigenetic control of T-helper-cell differentiation. *Nat. Rev. Immunol.* **2009**, *9*, 91–105. [[CrossRef](#)] [[PubMed](#)]
11. Tone, A.A.; Salvador, S.; Finlayson, S.J.; Tinker, A.V.; Kwon, J.S.; Lee, C.H.; Cohen, T.; Ehlen, T.; Lee, M.; Carey, M.S.; et al. The role of the fallopian tube in ovarian cancer. *Clin. Adv. Hematol. Oncol.* **2012**, *10*, 296–306. [[PubMed](#)]
12. Petrovska, M.; Dimitrov, D.G.; Michael, S.D. Quantitative changes in macrophage distribution in normal mouse ovary over the course of the estrous cycle examined with an image analysis system. *Am. J. Reprod. Immunol.* **1996**, *36*, 175–183. [[CrossRef](#)] [[PubMed](#)]
13. Takaya, R.; Fukaya, T.; Sasano, H.; Suzuki, T.; Tamura, M.; Yajima, A. Macrophages in normal cycling human ovaries; immunohistochemical localization and characterization. *Hum. Reprod.* **1997**, *12*, 1508–1512. [[CrossRef](#)] [[PubMed](#)]
14. Wu, R.; Van der Hoek, K.H.; Ryan, N.K.; Norman, R.J.; Robker, R.L. Macrophage contributions to ovarian function. *Hum. Reprod. Update* **2004**, *10*, 119–133. [[CrossRef](#)] [[PubMed](#)]



15. Tingen, C.M.; Kiesewetter, S.E.; Jozefik, J.; Thomas, C.; Tagler, D.; Shea, L.; Woodruff, T.K. A macrophage and theca cell-enriched stromal cell population influences growth and survival of immature murine follicles in vitro. *Reproduction* **2011**, *141*, 809–820. [[CrossRef](#)] [[PubMed](#)]
16. Lau, A.; Kollara, A.; St John, E.; Tone, A.A.; Virtanen, C.; Greenblatt, E.M.; King, W.A.; Brown, T.J. Altered expression of inflammation-associated genes in oviductal cells following follicular fluid exposure: Implications for ovarian carcinogenesis. *Exp. Biol. Med.* **2014**, *239*, 24–32. [[CrossRef](#)] [[PubMed](#)]
17. King, S.M.; Hilliard, T.S.; Wu, L.Y.; Jaffe, R.C.; Fazleabas, A.T.; Burdette, J.E. The impact of ovulation on fallopian tube epithelial cells: Evaluating three hypotheses connecting ovulation and serous ovarian cancer. *Endocr. Relat. Cancer* **2011**, *18*, 627–642. [[CrossRef](#)] [[PubMed](#)]
18. Johnson, P.A.; Giles, J.R. The hen as a model of ovarian cancer. *Nat. Rev. Cancer* **2013**, *13*, 432–436. [[CrossRef](#)] [[PubMed](#)]
19. Gong, T.T.; Wu, Q.J.; Vogtmann, E.; Lin, B.; Wang, Y.L. Age at menarche and risk of ovarian cancer: A meta-analysis of epidemiological studies. *Int. J. Cancer* **2013**, *132*, 2894–2900. [[CrossRef](#)] [[PubMed](#)]
20. Chiaffarino, F.; Pelucchi, C.; Parazzini, F.; Negri, E.; Franceschi, S.; Talamini, R.; Conti, E.; Montella, M.; La Vecchia, C. Reproductive and hormonal factors and ovarian cancer. *Ann. Oncol.* **2001**, *12*, 337–341. [[CrossRef](#)] [[PubMed](#)]
21. Fortner, R.T.; Ose, J.; Merritt, M.A.; Schock, H.; Tjonneland, A.; Hansen, L.; Overvad, K.; Dossus, L.; Clavel-Chapelon, F.; Baglietto, L.; et al. Reproductive and hormone-related risk factors for epithelial ovarian cancer by histologic pathways, invasiveness and histologic subtypes: Results from the EPIC cohort. *Int. J. Cancer* **2015**, *137*, 1196–1208. [[CrossRef](#)] [[PubMed](#)]
22. Espey, L.L. Ovulation as an inflammatory reaction—A hypothesis. *Biol. Reprod.* **1980**, *22*, 73–106. [[CrossRef](#)] [[PubMed](#)]
23. Machelon, V.; Emilie, D. Production of ovarian cytokines and their role in ovulation in the mammalian ovary. *Eur. Cytokine Netw.* **1997**, *8*, 137–143. [[PubMed](#)]
24. Chumduri, C.; Gurumurthy, R.K.; Zadora, P.K.; Mi, Y.; Meyer, T.F. Chlamydia infection promotes host DNA damage and proliferation but impairs the DNA damage response. *Cell. Host Microbe* **2013**, *13*, 746–758. [[CrossRef](#)] [[PubMed](#)]
25. Ingerslev, K.; Hogdall, E.; Schnack, T.H.; Skovrider-Ruminski, W.; Hogdall, C.; Blaakaer, J. The potential role of infectious agents and pelvic inflammatory disease in ovarian carcinogenesis. *Infect. Agent Cancer* **2017**, *12*, 25. [[CrossRef](#)] [[PubMed](#)]
26. Burney, R.O.; Giudice, L.C. Pathogenesis and pathophysiology of endometriosis. *Fertil. Steril.* **2012**, *98*, 511–519. [[CrossRef](#)] [[PubMed](#)]
27. Vercellini, P.; Crosignani, P.; Somigliana, E.; Viganò, P.; Buggio, L.; Bolis, G.; Fedele, L. The ‘incessant menstruation’ hypothesis: A mechanistic ovarian cancer model with implications for prevention. *Hum. Reprod.* **2011**, *26*, 2262–2273. [[CrossRef](#)] [[PubMed](#)]
28. Burghaus, S.; Haberle, L.; Schrauder, M.G.; Heusinger, K.; Thiel, F.C.; Hein, A.; Wachter, D.; Strehl, J.; Hartmann, A.; Ekici, A.B.; et al. Endometriosis as a risk factor for ovarian or endometrial cancer—Results of a hospital-based case-control study. *BMC Cancer* **2015**, *15*, 751. [[CrossRef](#)] [[PubMed](#)]
29. Sayasneh, A.; Tsivos, D.; Crawford, R. Endometriosis and ovarian cancer: A systematic review. *ISRN Obstet. Gynecol.* **2011**, *2011*, 140310. [[CrossRef](#)] [[PubMed](#)]
30. Rizi, B.S.; Nagrath, D. Linking omentum and ovarian cancer: NO. *Oncoscience* **2015**, *2*, 797–798. [[PubMed](#)]
31. Gunderson, C.C.; Ding, K.; Dvorak, J.; Moore, K.N.; McMeekin, D.S.; Benbrook, D.M. The pro-inflammatory effect of obesity on high grade serous ovarian cancer. *Gynecol. Oncol.* **2016**, *143*, 40–45. [[CrossRef](#)] [[PubMed](#)]
32. Poole, E.M.; Lee, I.M.; Ridker, P.M.; Buring, J.E.; Hankinson, S.E.; Tworoger, S.S. A prospective study of circulating C-reactive protein, interleukin-6, and tumor necrosis factor alpha receptor 2 levels and risk of ovarian cancer. *Am. J. Epidemiol.* **2013**, *178*, 1256–1264. [[CrossRef](#)] [[PubMed](#)]
33. Ose, J.; Schock, H.; Tjonneland, A.; Hansen, L.; Overvad, K.; Dossus, L.; Clavel-Chapelon, F.; Baglietto, L.; Boeing, H.; Trichopolou, A.; et al. Inflammatory Markers and Risk of Epithelial Ovarian Cancer by Tumor Subtypes: The EPIC Cohort. *Cancer Epidemiol. Biomark. Prev.* **2015**, *24*, 951–961. [[CrossRef](#)] [[PubMed](#)]
34. Duleba, A.J.; Dokras, A. Is PCOS an inflammatory process? *Fertil. Steril.* **2012**, *97*, 7–12. [[CrossRef](#)] [[PubMed](#)]
35. Kelly, C.C.; Lyall, H.; Petrie, J.R.; Gould, G.W.; Connell, J.M.; Sattar, N. Low grade chronic inflammation in women with polycystic ovarian syndrome. *J. Clin. Endocrinol. Metab.* **2001**, *86*, 2453–2455. [[CrossRef](#)] [[PubMed](#)]

36. Escobar-Morreale, H.F.; Luque-Ramirez, M.; Gonzalez, F. Circulating inflammatory markers in polycystic ovary syndrome: A systematic review and metaanalysis. *Fertil. Steril.* **2011**, *95*, 1048–1058. [\[CrossRef\]](#) [\[PubMed\]](#)
37. Gonzalez, F.; Rote, N.S.; Minium, J.; Kirwan, J.P. Evidence of proatherogenic inflammation in polycystic ovary syndrome. *Metabolism* **2009**, *58*, 954–962. [\[CrossRef\]](#) [\[PubMed\]](#)
38. Glintborg, D.; Andersen, M.; Richelsen, B.; Bruun, J.M. Plasma monocyte chemoattractant protein-1 (MCP-1) and macrophage inflammatory protein-1 $\alpha$  are increased in patients with polycystic ovary syndrome (PCOS) and associated with adiposity, but unaffected by pioglitazone treatment. *Clin. Endocrinol.* **2009**, *71*, 652–658. [\[CrossRef\]](#) [\[PubMed\]](#)
39. Escobar-Morreale, H.F.; Botella-Carretero, J.I.; Villuendas, G.; Sancho, J.; San Millan, J.L. Serum interleukin-18 concentrations are increased in the polycystic ovary syndrome: Relationship to insulin resistance and to obesity. *J. Clin. Endocrinol. Metab.* **2004**, *89*, 806–811. [\[CrossRef\]](#) [\[PubMed\]](#)
40. Yang, Y.; Qiao, J.; Li, R.; Li, M.Z. Is interleukin-18 associated with polycystic ovary syndrome? *Reprod. Biol. Endocrinol.* **2011**, *9*, 7. [\[CrossRef\]](#) [\[PubMed\]](#)
41. Tarkun, I.; Cetinarslan, B.; Turemen, E.; Canturk, Z.; Biyikli, M. Association between Circulating Tumor Necrosis Factor- $\alpha$ , Interleukin-6, and Insulin Resistance in Normal-Weight Women with Polycystic Ovary Syndrome. *Metab. Syndr. Relat. Disord.* **2006**, *4*, 122–128. [\[CrossRef\]](#) [\[PubMed\]](#)
42. Vgontzas, A.N.; Trakada, G.; Bixler, E.O.; Lin, H.M.; Pejovic, S.; Zoumakis, E.; Chrousos, G.P.; Legro, R.S. Plasma interleukin 6 levels are elevated in polycystic ovary syndrome independently of obesity or sleep apnea. *Metabolism* **2006**, *55*, 1076–1082. [\[CrossRef\]](#) [\[PubMed\]](#)
43. Dinger, Y.; Akcay, T.; Erdem, T.; Ilker Saygili, E.; Gundogdu, S. DNA damage, DNA susceptibility to oxidation and glutathione level in women with polycystic ovary syndrome. *Scand. J. Clin. Lab. Investig.* **2005**, *65*, 721–728.
44. Harris, H.R.; Terry, K.L. Polycystic ovary syndrome and risk of endometrial, ovarian, and breast cancer: A systematic review. *Fertil. Res. Pract.* **2016**, *2*, 14. [\[CrossRef\]](#) [\[PubMed\]](#)
45. Heller, D.S.; Westhoff, C.; Gordon, R.E.; Katz, N. The relationship between perineal cosmetic talc usage and ovarian talc particle burden. *Am. J. Obstet. Gynecol.* **1996**, *174*, 1507–1510. [\[CrossRef\]](#)
46. Henderson, W.J.; Hamilton, T.C.; Griffiths, K. Talc in normal and malignant ovarian tissue. *Lancet* **1979**, *1*, 499. [\[CrossRef\]](#)
47. Muscat, J.E.; Huncharek, M.S. Perineal talc use and ovarian cancer: A critical review. *Eur. J. Cancer Prev.* **2008**, *17*, 139–146. [\[CrossRef\]](#) [\[PubMed\]](#)
48. Brasky, T.M.; Moysich, K.B.; Cohn, D.E.; White, E. Non-steroidal anti-inflammatory drugs and endometrial cancer risk in the VITamins And Lifestyle (VITAL) cohort. *Gynecol. Oncol.* **2013**, *128*, 113–119. [\[CrossRef\]](#) [\[PubMed\]](#)
49. Prizment, A.E.; Folsom, A.R.; Anderson, K.E. Nonsteroidal anti-inflammatory drugs and risk for ovarian and endometrial cancers in the Iowa Women's Health Study. *Cancer Epidemiol. Biomark. Prev.* **2010**, *19*, 435–442. [\[CrossRef\]](#) [\[PubMed\]](#)
50. Fairfield, K.M.; Hunter, D.J.; Fuchs, C.S.; Colditz, G.A.; Hankinson, S.E. Aspirin, other NSAIDs, and ovarian cancer risk (United States). *Cancer Causes Control.* **2002**, *13*, 535–542. [\[CrossRef\]](#) [\[PubMed\]](#)
51. Trabert, B.; Poole, E.M.; White, E.; Visvanathan, K.; Adami, H.O.; Anderson, G.L.; Brasky, T.M.; Brinton, L.A.; Fortner, R.T.; Gaudet, M.; et al. Analgesic Use and Ovarian Cancer Risk: An Analysis in the Ovarian Cancer Cohort Consortium. *J. Natl. Cancer Inst.* **2018**. [\[CrossRef\]](#) [\[PubMed\]](#)
52. Peres, L.C.; Camacho, F.; Abbott, S.E.; Alberg, A.J.; Bandera, E.V.; Barnholtz-Sloan, J.; Bondy, M.; Cote, M.L.; Crankshaw, S.; Funkhouser, E.; et al. Analgesic medication use and risk of epithelial ovarian cancer in African American women. *Br. J. Cancer* **2016**, *114*, 819–825. [\[CrossRef\]](#) [\[PubMed\]](#)
53. Rodriguez-Burford, C.; Barnes, M.N.; Oelschlager, D.K.; Myers, R.B.; Talley, L.I.; Partridge, E.E.; Grizzle, W.E. Effects of nonsteroidal anti-inflammatory agents (NSAIDs) on ovarian carcinoma cell lines: Preclinical evaluation of NSAIDs as chemopreventive agents. *Clin. Cancer Res.* **2002**, *8*, 202–209. [\[PubMed\]](#)
54. Arango, H.A.; Icely, S.; Roberts, W.S.; Cavanagh, D.; Becker, J.L. Aspirin effects on endometrial cancer cell growth. *Obstet. Gynecol.* **2001**, *97*, 423–427. [\[PubMed\]](#)
55. Gao, J.; Niwa, K.; Sun, W.; Takemura, M.; Lian, Z.; Onogi, K.; Seishima, M.; Mori, H.; Tamaya, T. Non-steroidal anti-inflammatory drugs inhibit cellular proliferation and upregulate cyclooxygenase-2 protein expression in endometrial cancer cells. *Cancer Sci.* **2004**, *95*, 901–907. [\[CrossRef\]](#) [\[PubMed\]](#)

56. Li, H.L.; Zhang, H.W.; Chen, D.D.; Zhong, L.; Ren, X.D.; St-Tu, R. JTE-522, a selective COX-2 inhibitor, inhibits cell proliferation and induces apoptosis in RL95-2 cells. *Acta Pharmacol Sin.* **2002**, *23*, 631–637. [[PubMed](#)]
57. Hsu, C.S.; Li, Y. Aspirin potently inhibits oxidative DNA strand breaks: Implications for cancer chemoprevention. *Biochem. Biophys. Res. Commun.* **2002**, *293*, 705–709. [[PubMed](#)]
58. Hales, D.B.; Zhuge, Y.; Lagman, J.A.; Ansenberger, K.; Mahon, C.; Barua, A.; Luborsky, J.L.; Bahr, J.M. Cyclooxygenases expression and distribution in the normal ovary and their role in ovarian cancer in the domestic hen (*Gallus domesticus*). *Endocrine* **2008**, *33*, 235–244. [[CrossRef](#)] [[PubMed](#)]
59. Urick, M.E.; Giles, J.R.; Johnson, P.A. VEGF expression and the effect of NSAIDs on ascites cell proliferation in the hen model of ovarian cancer. *Gynecol. Oncol.* **2008**, *110*, 418–424. [[CrossRef](#)] [[PubMed](#)]
60. Urick, M.E.; Giles, J.R.; Johnson, P.A. Dietary aspirin decreases the stage of ovarian cancer in the hen. *Gynecol. Oncol.* **2009**, *112*, 166–170. [[CrossRef](#)] [[PubMed](#)]
61. Hanahan, D.; Weinberg, R.A. Hallmarks of cancer: The next generation. *Cell* **2011**, *144*, 646–674. [[CrossRef](#)] [[PubMed](#)]
62. Cancer Genome Atlas Research, N. Integrated genomic analyses of ovarian carcinoma. *Nature* **2011**, *474*, 609–615.
63. Toss, A.; Tomasello, C.; Razzaboni, E.; Contu, G.; Grandi, G.; Cagnacci, A.; Schilder, R.J.; Cortesi, L. Hereditary ovarian cancer: Not only BRCA 1 and 2 genes. *Biomed. Res. Int.* **2015**, *2015*, 341723. [[CrossRef](#)] [[PubMed](#)]
64. Liu, J.; Yang, G.; Thompson-Lanza, J.A.; Glassman, A.; Hayes, K.; Patterson, A.; Marquez, R.T.; Auersperg, N.; Yu, Y.; Hahn, W.C.; et al. A genetically defined model for human ovarian cancer. *Cancer Res.* **2004**, *64*, 1655–1663. [[CrossRef](#)] [[PubMed](#)]
65. Szotek, P.P.; Pieretti-Vanmarcke, R.; Masiakos, P.T.; Dinulescu, D.M.; Connolly, D.; Foster, R.; Dombkowski, D.; Preffer, F.; Maclaughlin, D.T.; Donahoe, P.K. Ovarian cancer side population defines cells with stem cell-like characteristics and Mullerian Inhibiting Substance responsiveness. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 11154–11159. [[CrossRef](#)] [[PubMed](#)]
66. Zhang, S.; Balch, C.; Chan, M.W.; Lai, H.C.; Matei, D.; Schilder, J.M.; Yan, P.S.; Huang, T.H.; Nephew, K.P. Identification and characterization of ovarian cancer-initiating cells from primary human tumors. *Cancer Res.* **2008**, *68*, 4311–4320. [[CrossRef](#)] [[PubMed](#)]
67. Lane, D.; Matte, I.; Rancourt, C.; Piche, A. Prognostic significance of IL-6 and IL-8 ascites levels in ovarian cancer patients. *BMC Cancer* **2011**, *11*, 210. [[CrossRef](#)] [[PubMed](#)]
68. Masoumi-Moghaddam, S.; Amini, A.; Wei, A.Q.; Robertson, G.; Morris, D.L. Intratumoral interleukin-6 predicts ascites formation in patients with epithelial ovarian cancer: A potential tool for close monitoring. *J. Ovarian Res.* **2015**, *8*, 58. [[CrossRef](#)] [[PubMed](#)]
69. Rath, K.S.; Funk, H.M.; Bowling, M.C.; Richards, W.E.; Drew, A.F. Expression of soluble interleukin-6 receptor in malignant ovarian tissue. *Am. J. Obstet. Gynecol.* **2010**, *203*, 230.e1–230.e8. [[CrossRef](#)] [[PubMed](#)]
70. Lo, C.W.; Chen, M.W.; Hsiao, M.; Wang, S.; Chen, C.A.; Hsiao, S.M.; Chang, J.S.; Lai, T.C.; Rose-John, S.; Kuo, M.L.; et al. IL-6 trans-signaling in formation and progression of malignant ascites in ovarian cancer. *Cancer Res.* **2011**, *71*, 424–434. [[CrossRef](#)] [[PubMed](#)]
71. Dalal, V.; Kumar, R.; Kumar, S.; Sharma, A.; Kumar, L.; Sharma, J.B.; Roy, K.K.; Singh, N.; Vanamail, P. Biomarker potential of IL-6 and VEGF-A in ascitic fluid of epithelial ovarian cancer patients. *Clin. Chim. Acta* **2018**, *482*, 27–32. [[CrossRef](#)] [[PubMed](#)]
72. Bapat, S.A.; Mali, A.M.; Koppikar, C.B.; Kurrey, N.K. Stem and progenitor-like cells contribute to the aggressive behavior of human epithelial ovarian cancer. *Cancer Res.* **2005**, *65*, 3025–3029. [[CrossRef](#)] [[PubMed](#)]
73. Wang, D.; Xiang, T.; Zhao, Z.; Lin, K.; Yin, P.; Jiang, L.; Liang, Z.; Zhu, B. Autocrine interleukin-23 promotes self-renewal of CD133+ ovarian cancer stem-like cells. *Oncotarget* **2016**, *7*, 76006–76020. [[CrossRef](#)] [[PubMed](#)]
74. Maccio, A.; Madeddu, C. Inflammation and ovarian cancer. *Cytokine* **2012**, *58*, 133–147. [[CrossRef](#)] [[PubMed](#)]
75. Agarwal, A.; Gupta, S.; Sharma, R.K. Role of oxidative stress in female reproduction. *Reprod. Biol. Endocrinol.* **2005**, *3*, 28. [[CrossRef](#)] [[PubMed](#)]
76. Shkolnik, K.; Tadmor, A.; Ben-Dor, S.; Nevo, N.; Galiani, D.; Dekel, N. Reactive oxygen species are indispensable in ovulation. *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 1462–1467. [[CrossRef](#)] [[PubMed](#)]
77. Waris, G.; Ahsan, H. Reactive oxygen species: Role in the development of cancer and various chronic conditions. *J. Carcinog.* **2006**, *5*, 14. [[CrossRef](#)] [[PubMed](#)]
78. Liou, G.Y.; Storz, P. Reactive oxygen species in cancer. *Free Radic. Res.* **2010**, *44*, 479–496. [[CrossRef](#)] [[PubMed](#)]

79. King, S.M.; Quartuccio, S.M.; Vanderhyden, B.C.; Burdette, J.E. Early transformative changes in normal ovarian surface epithelium induced by oxidative stress require Akt upregulation, DNA damage and epithelial-stromal interaction. *Carcinogenesis* **2013**, *34*, 1125–1133. [[CrossRef](#)] [[PubMed](#)]
80. Levanon, K.; Ng, V.; Piao, H.Y.; Zhang, Y.; Chang, M.C.; Roh, M.H.; Kindelberger, D.W.; Hirsch, M.S.; Crum, C.P.; Marto, J.A.; et al. Primary ex vivo cultures of human fallopian tube epithelium as a model for serous ovarian carcinogenesis. *Oncogene* **2010**, *29*, 1103–1113. [[CrossRef](#)] [[PubMed](#)]
81. Huang, H.S.; Chu, S.C.; Hsu, C.F.; Chen, P.C.; Ding, D.C.; Chang, M.Y.; Chu, T.Y. Mutagenic, surviving and tumorigenic effects of follicular fluid in the context of p53 loss: Initiation of fimbria carcinogenesis. *Carcinogenesis* **2015**, *36*, 1419–1428. [[CrossRef](#)] [[PubMed](#)]
82. Kalinina, E.V.; Chernov, N.N.; Saprin, A.N.; Kotova, Y.N.; Gavrilova, Y.A.; Chermnykh, N.S.; Shcherbak, N.P. Expression of genes for thioredoxin 1 and thioredoxin 2 in multidrug resistance ovarian carcinoma cells SKVLB. *Bull. Exp. Biol. Med.* **2007**, *144*, 301–303. [[CrossRef](#)] [[PubMed](#)]
83. Ohno, T.; Hirota, K.; Nakamura, H.; Masutani, H.; Sasada, T.; Yodoi, J. Thioredoxin and Its Involvement in the Redox Regulation of Transcription Factors, NF- $\kappa$ B and AP-1. In *Oxygen Homeostasis and Its Dynamics*; Springer: Tokyo, Japan, 1998; pp. 450–456.
84. Van der Wijst, M.G.; Huisman, C.; Mposhi, A.; Roelfes, G.; Rots, M.G. Targeting Nrf2 in healthy and malignant ovarian epithelial cells: Protection versus promotion. *Mol. Oncol.* **2015**, *9*, 1259–1273. [[CrossRef](#)] [[PubMed](#)]
85. Klotz, L.O.; Sanchez-Ramos, C.; Prieto-Arroyo, I.; Urbanek, P.; Steinbrenner, H.; Monsalve, M. Redox regulation of FoxO transcription factors. *Redox Biol* **2015**, *6*, 51–72. [[CrossRef](#)] [[PubMed](#)]
86. Ozdemir, F.; Altinisik, J.; Karateke, A.; Coksuer, H.; Buyru, N. Methylation of tumor suppressor genes in ovarian cancer. *Exp. Ther. Med.* **2012**, *4*, 1092–1096. [[CrossRef](#)] [[PubMed](#)]
87. Niwa, T.; Tsukamoto, T.; Toyoda, T.; Mori, A.; Tanaka, H.; Maekita, T.; Ichinose, M.; Tatematsu, M.; Ushijima, T. Inflammatory processes triggered by Helicobacter pylori infection cause aberrant DNA methylation in gastric epithelial cells. *Cancer Res.* **2010**, *70*, 1430–1440. [[CrossRef](#)] [[PubMed](#)]
88. Ding, N.; Bonham, E.M.; Hannon, B.E.; Amick, T.R.; Baylin, S.B.; O'Hagan, H.M. Mismatch repair proteins recruit DNA methyltransferase 1 to sites of oxidative DNA damage. *J. Mol. Cell. Biol.* **2016**, *8*, 244–254. [[CrossRef](#)] [[PubMed](#)]
89. Maiuri, A.R.; Peng, M.; Sriramkumar, S.; Kamplain, C.M.; DeStefano Shields, C.E.; Sears, C.L.; O'Hagan, H.M. Mismatch Repair Proteins Initiate Epigenetic Alterations during Inflammation-Driven Tumorigenesis. *Cancer Res.* **2017**, *77*, 3467–3478. [[CrossRef](#)] [[PubMed](#)]
90. Sapoznik, S.; Bahar-Shany, K.; Brand, H.; Pinto, Y.; Gabay, O.; Glick-Saar, E.; Dor, C.; Zadok, O.; Barshack, I.; Zundevich, A.; et al. Activation-Induced Cytidine Deaminase Links Ovulation-Induced Inflammation and Serous Carcinogenesis. *Neoplasia* **2016**, *18*, 90–99. [[CrossRef](#)] [[PubMed](#)]
91. Gupta, M.; Babic, A.; Beck, A.H.; Terry, K. TNF- $\alpha$  expression, risk factors, and inflammatory exposures in ovarian cancer: Evidence for an inflammatory pathway of ovarian carcinogenesis? *Hum. Pathol.* **2016**, *54*, 82–91. [[CrossRef](#)] [[PubMed](#)]
92. Moradi, M.M.; Carson, L.F.; Weinberg, B.; Haney, A.F.; Twigg, L.B.; Ramakrishnan, S. Serum and ascitic fluid levels of interleukin-1, interleukin-6, and tumor necrosis factor- $\alpha$  in patients with ovarian epithelial cancer. *Cancer* **1993**, *72*, 2433–2440. [[CrossRef](#)]
93. Szlosarek, P.W.; Grimshaw, M.J.; Kulbe, H.; Wilson, J.L.; Wilbanks, G.D.; Burke, F.; Balkwill, F.R. Expression and regulation of tumor necrosis factor alpha in normal and malignant ovarian epithelium. *Mol. Cancer Ther.* **2006**, *5*, 382–390. [[CrossRef](#)] [[PubMed](#)]
94. Kulbe, H.; Thompson, R.; Wilson, J.L.; Robinson, S.; Hagemann, T.; Fatah, R.; Gould, D.; Ayhan, A.; Balkwill, F. The inflammatory cytokine tumor necrosis factor- $\alpha$  generates an autocrine tumor-promoting network in epithelial ovarian cancer cells. *Cancer Res.* **2007**, *67*, 585–592. [[CrossRef](#)] [[PubMed](#)]
95. Charles, K.A.; Kulbe, H.; Soper, R.; Escorcio-Correia, M.; Lawrence, T.; Schultheis, A.; Chakravarty, P.; Thompson, R.G.; Kollias, G.; Smyth, J.F.; et al. The tumor-promoting actions of TNF- $\alpha$  involve TNFR1 and IL-17 in ovarian cancer in mice and humans. *J. Clin. Investig.* **2009**, *119*, 3011–3023. [[CrossRef](#)] [[PubMed](#)]
96. Berek, J.S.; Chung, C.; Kaldi, K.; Watson, J.M.; Knox, R.M.; Martinez-Maza, O. Serum interleukin-6 levels correlate with disease status in patients with epithelial ovarian cancer. *Am. J. Obstet. Gynecol.* **1991**, *164*, 1038–1042; discussion 1042–1033. [[CrossRef](#)]



97. Scambia, G.; Testa, U.; Benedetti Panici, P.; Foti, E.; Martucci, R.; Gadducci, A.; Perillo, A.; Facchini, V.; Peschle, C.; Mancuso, S. Prognostic significance of interleukin 6 serum levels in patients with ovarian cancer. *Br. J. Cancer* **1995**, *71*, 354. [[CrossRef](#)] [[PubMed](#)]
98. Alberti, C.; Pinciroli, P.; Valeri, B.; Ferri, R.; Ditto, A.; Umezawa, K.; Sensi, M.; Canevari, S.; Tomassetti, A. Ligand-dependent EGFR activation induces the co-expression of IL-6 and PAI-1 via the NFkB pathway in advanced-stage epithelial ovarian cancer. *Oncogene* **2012**, *31*, 4139–4149. [[CrossRef](#)] [[PubMed](#)]
99. Hagemann, T.; Robinson, S.C.; Thompson, R.G.; Charles, K.; Kulbe, H.; Balkwill, F.R. Ovarian cancer cell-derived migration inhibitory factor enhances tumor growth, progression, and angiogenesis. *Mol. Cancer Ther.* **2007**, *6*, 1993–2002. [[CrossRef](#)] [[PubMed](#)]
100. Oh, K.; Moon, H.G.; Lee, D.S.; Yoo, Y.B. Tissue transglutaminase-interleukin-6 axis facilitates peritoneal tumor spreading and metastasis of human ovarian cancer cells. *Lab. Anim. Res.* **2015**, *31*, 188–197. [[CrossRef](#)] [[PubMed](#)]
101. Wang, Y.; Li, L.; Guo, X.; Jin, X.; Sun, W.; Zhang, X.; Xu, R.C. Interleukin-6 signaling regulates anchorage-independent growth, proliferation, adhesion and invasion in human ovarian cancer cells. *Cytokine* **2012**, *59*, 228–236. [[CrossRef](#)] [[PubMed](#)]
102. Takaishi, K.; Komohara, Y.; Tashiro, H.; Ohtake, H.; Nakagawa, T.; Katabuchi, H.; Takeya, M. Involvement of M2-polarized macrophages in the ascites from advanced epithelial ovarian carcinoma in tumor progression via Stat3 activation. *Cancer Sci.* **2010**, *101*, 2128–2136. [[CrossRef](#)] [[PubMed](#)]
103. Maccio, A.; Lai, P.; Santona, M.C.; Pagliara, L.; Melis, G.B.; Mantovani, G. High serum levels of soluble IL-2 receptor, cytokines, and C reactive protein correlate with impairment of T cell response in patients with advanced epithelial ovarian cancer. *Gynecol. Oncol.* **1998**, *69*, 248–252. [[CrossRef](#)] [[PubMed](#)]
104. Rabinovich, A.; Medina, L.; Piura, B.; Segal, S.; Huleihel, M. Regulation of ovarian carcinoma SKOV-3 cell proliferation and secretion of MMPs by autocrine IL-6. *Anticancer Res.* **2007**, *27*, 267–272. [[PubMed](#)]
105. Runesson, E.; Bostrom, E.K.; Janson, P.O.; Brannstrom, M. The human preovulatory follicle is a source of the chemotactic cytokine interleukin-8. *Mol. Hum. Reprod* **1996**, *2*, 245–250. [[CrossRef](#)] [[PubMed](#)]
106. Brannstrom, M.; Mayrhofer, G.; Robertson, S.A. Localization of leukocyte subsets in the rat ovary during the periovulatory period. *Biol. Reprod.* **1993**, *48*, 277–286. [[CrossRef](#)] [[PubMed](#)]
107. Kassim, S.K.; El-Salahy, E.M.; Fayed, S.T.; Helal, S.A.; Helal, T.; Azzam Eel, D.; Khalifa, A. Vascular endothelial growth factor and interleukin-8 are associated with poor prognosis in epithelial ovarian cancer patients. *Clin. Biochem.* **2004**, *37*, 363–369. [[CrossRef](#)] [[PubMed](#)]
108. Ivarsson, K.; Runesson, E.; Sundfeldt, K.; Haeger, M.; Hedin, L.; Janson, P.O.; Brannstrom, M. The chemotactic cytokine interleukin-8—A cyst fluid marker for malignant epithelial ovarian cancer? *Gynecol. Oncol.* **1998**, *71*, 420–423. [[CrossRef](#)] [[PubMed](#)]
109. Radke, J.; Schmidt, D.; Bohme, M.; Schmidt, U.; Weise, W.; Morenz, J. Cytokine level in malignant ascites and peripheral blood of patients with advanced ovarian carcinoma. *Geburtshilfe Frauenheilkd.* **1996**, *56*, 83–87. [[CrossRef](#)] [[PubMed](#)]
110. Wang, Y.; Xu, R.C.; Zhang, X.L.; Niu, X.L.; Qu, Y.; Li, L.Z.; Meng, X.Y. Interleukin-8 secretion by ovarian cancer cells increases anchorage-independent growth, proliferation, angiogenic potential, adhesion and invasion. *Cytokine* **2012**, *59*, 145–155. [[CrossRef](#)] [[PubMed](#)]
111. Hirose, K.; Hakozaiki, M.; Nyunoya, Y.; Kobayashi, Y.; Matsushita, K.; Takenouchi, T.; Mikata, A.; Mukaida, N.; Matsushima, K. Chemokine gene transfection into tumour cells reduced tumorigenicity in nude mice in association with neutrophilic infiltration. *Br. J. Cancer* **1995**, *72*, 708–714. [[CrossRef](#)] [[PubMed](#)]
112. Lee, L.F.; Hellendall, R.P.; Wang, Y.; Haskill, J.S.; Mukaida, N.; Matsushima, K.; Ting, J.P. IL-8 reduced tumorigenicity of human ovarian cancer in vivo due to neutrophil infiltration. *J. Immunol.* **2000**, *164*, 2769–2775. [[CrossRef](#)] [[PubMed](#)]
113. Tokumura, A.; Miyake, M.; Nishioka, Y.; Yamano, S.; Aono, T.; Fukuzawa, K. Production of lysophosphatidic acids by lysophospholipase D in human follicular fluids of In vitro fertilization patients. *Biol. Reprod.* **1999**, *61*, 195–199. [[CrossRef](#)] [[PubMed](#)]
114. Chen, S.U.; Chou, C.H.; Lee, H.; Ho, C.H.; Lin, C.W.; Yang, Y.S. Lysophosphatidic acid up-regulates expression of interleukin-8 and -6 in granulosa-lutein cells through its receptors and nuclear factor-κB dependent pathways: Implications for angiogenesis of corpus luteum and ovarian hyperstimulation syndrome. *J. Clin. Endocrinol. Metab.* **2008**, *93*, 935–943. [[CrossRef](#)] [[PubMed](#)]



115. Fang, X.; Gaudette, D.; Furui, T.; Mao, M.; Estrella, V.; Eder, A.; Pustilnik, T.; Sasagawa, T.; Lapushin, R.; Yu, S.; et al. Lysophospholipid growth factors in the initiation, progression, metastases, and management of ovarian cancer. *Ann. N. Y. Acad. Sci.* **2000**, *905*, 188–208. [[CrossRef](#)] [[PubMed](#)]
116. Xu, Y.; Shen, Z.; Wiper, D.W.; Wu, M.; Morton, R.E.; Elson, P.; Kennedy, A.W.; Belinson, J.; Markman, M.; Casey, G. Lysophosphatidic acid as a potential biomarker for ovarian and other gynecologic cancers. *JAMA* **1998**, *280*, 719–723. [[CrossRef](#)] [[PubMed](#)]
117. Xiao, Y.J.; Schwartz, B.; Washington, M.; Kennedy, A.; Webster, K.; Belinson, J.; Xu, Y. Electrospray ionization mass spectrometry analysis of lysophospholipids in human ascitic fluids: Comparison of the lysophospholipid contents in malignant vs nonmalignant ascitic fluids. *Anal. Biochem.* **2001**, *290*, 302–313. [[CrossRef](#)] [[PubMed](#)]
118. Mills, G.B.; Eder, A.; Fang, X.; Hasegawa, Y.; Mao, M.; Lu, Y.; Tanyi, J.; Tabassam, F.H.; Wiener, J.; Lapushin, R.; et al. Critical role of lysophospholipids in the pathophysiology, diagnosis, and management of ovarian cancer. *Cancer Treat. Res.* **2002**, *107*, 259–283. [[PubMed](#)]
119. Westermann, A.M.; Havik, E.; Postma, F.R.; Beijnen, J.H.; Dalesio, O.; Moolenaar, W.H.; Rodenhuis, S. Malignant effusions contain lysophosphatidic acid (LPA)-like activity. *Ann. Oncol.* **1998**, *9*, 437–442. [[CrossRef](#)] [[PubMed](#)]
120. Xu, Y.; Gaudette, D.C.; Boynton, J.D.; Frankel, A.; Fang, X.J.; Sharma, A.; Hurteau, J.; Casey, G.; Goodbody, A.; Mellors, A.; et al. Characterization of an ovarian cancer activating factor in ascites from ovarian cancer patients. *Clin. Cancer Res.* **1995**, *1*, 1223–1232. [[PubMed](#)]
121. Symowicz, J.; Adley, B.P.; Woo, M.M.; Auersperg, N.; Hudson, L.G.; Stack, M.S. Cyclooxygenase-2 functions as a downstream mediator of lysophosphatidic acid to promote aggressive behavior in ovarian carcinoma cells. *Cancer Res.* **2005**, *65*, 2234–2242. [[CrossRef](#)] [[PubMed](#)]
122. Fang, X.; Yu, S.; Bast, R.C.; Liu, S.; Xu, H.J.; Hu, S.X.; LaPushin, R.; Claret, F.X.; Aggarwal, B.B.; Lu, Y.; et al. Mechanisms for lysophosphatidic acid-induced cytokine production in ovarian cancer cells. *J. Biol. Chem.* **2004**, *279*, 9653–9661. [[CrossRef](#)] [[PubMed](#)]
123. Saunders, J.A.; Rogers, L.C.; Klomsiri, C.; Poole, L.B.; Daniel, L.W. Reactive oxygen species mediate lysophosphatidic acid induced signaling in ovarian cancer cells. *Free Radic. Biol. Med.* **2010**, *49*, 2058–2067. [[CrossRef](#)] [[PubMed](#)]
124. Lu, J.; Xiao Yj, Y.J.; Baudhuin, L.M.; Hong, G.; Xu, Y. Role of ether-linked lysophosphatidic acids in ovarian cancer cells. *J. Lipid Res.* **2002**, *43*, 463–476. [[PubMed](#)]
125. Ali-Fehmi, R.; Morris, R.T.; Bandyopadhyay, S.; Che, M.; Schimp, V.; Malone, J.M., Jr.; Munkarah, A.R. Expression of cyclooxygenase-2 in advanced stage ovarian serous carcinoma: Correlation with tumor cell proliferation, apoptosis, angiogenesis, and survival. *Am. J. Obstet. Gynecol.* **2005**, *192*, 819–825. [[CrossRef](#)] [[PubMed](#)]
126. Rask, K.; Zhu, Y.; Wang, W.; Hedin, L.; Sundfeldt, K. Ovarian epithelial cancer: A role for PGE2-synthesis and signalling in malignant transformation and progression. *Mol. Cancer* **2006**, *5*, 62. [[CrossRef](#)] [[PubMed](#)]
127. Heinonen, P.K.; Metsä-Ketela, T. Prostaglandin and thromboxane production in ovarian cancer tissue. *Gynecol. Obstet. Investig.* **1984**, *18*, 225–229. [[CrossRef](#)] [[PubMed](#)]
128. Obermajer, N.; Muthuswamy, R.; Odunsi, K.; Edwards, R.P.; Kalinski, P. PGE(2)-induced CXCL12 production and CXCR4 expression controls the accumulation of human MDSCs in ovarian cancer environment. *Cancer Res.* **2011**, *71*, 7463–7470. [[CrossRef](#)] [[PubMed](#)]
129. Daikoku, T.; Tranguch, S.; Trofimova, I.N.; Dinulescu, D.M.; Jacks, T.; Nikitin, A.Y.; Connolly, D.C.; Dey, S.K. Cyclooxygenase-1 is overexpressed in multiple genetically engineered mouse models of epithelial ovarian cancer. *Cancer Res.* **2006**, *66*, 2527–2531. [[CrossRef](#)] [[PubMed](#)]
130. Li, W.; Cai, J.H.; Zhang, J.; Tang, Y.X.; Wan, L. Effects of cyclooxygenase inhibitors in combination with taxol on expression of cyclin D1 and Ki-67 in a xenograft model of ovarian carcinoma. *Int. J. Mol. Sci.* **2012**, *13*, 9741–9753. [[CrossRef](#)] [[PubMed](#)]
131. Paweletz, N.; Knierim, M. Tumor-related angiogenesis. *Crit. Rev. Oncol. Hematol.* **1989**, *9*, 197–242. [[CrossRef](#)]
132. Potente, M.; Gerhardt, H.; Carmeliet, P. Basic and therapeutic aspects of angiogenesis. *Cell* **2011**, *146*, 873–887. [[CrossRef](#)] [[PubMed](#)]
133. Burke, B.; Giannoudis, A.; Corke, K.P.; Gill, D.; Wells, M.; Ziegler-Heitbrock, L.; Lewis, C.E. Hypoxia-induced gene expression in human macrophages: Implications for ischemic tissues and hypoxia-regulated gene therapy. *Am. J. Pathol.* **2003**, *163*, 1233–1243. [[CrossRef](#)]

134. Murdoch, C.; Muthana, M.; Lewis, C.E. Hypoxia regulates macrophage functions in inflammation. *J. Immunol.* **2005**, *175*, 6257–6263. [\[CrossRef\]](#) [\[PubMed\]](#)
135. Ferrara, N.; Gerber, H.P.; LeCouter, J. The biology of VEGF and its receptors. *Nat. Med.* **2003**, *9*, 669–676. [\[CrossRef\]](#) [\[PubMed\]](#)
136. Wheeler-Jones, C.; Abu-Ghazaleh, R.; Cospedal, R.; Houliston, R.A.; Martin, J.; Zachary, I. Vascular endothelial growth factor stimulates prostacyclin production and activation of cytosolic phospholipase A2 in endothelial cells via p42/p44 mitogen-activated protein kinase. *FEBS Lett.* **1997**, *420*, 28–32. [\[CrossRef\]](#)
137. Gerber, H.P.; McMurtrey, A.; Kowalski, J.; Yan, M.; Keyt, B.A.; Dixit, V.; Ferrara, N. Vascular endothelial growth factor regulates endothelial cell survival through the phosphatidylinositol 3'-kinase/Akt signal transduction pathway. Requirement for Flk-1/KDR activation. *J. Biol. Chem.* **1998**, *273*, 30336–30343. [\[CrossRef\]](#) [\[PubMed\]](#)
138. Gerber, H.P.; Dixit, V.; Ferrara, N. Vascular endothelial growth factor induces expression of the antiapoptotic proteins Bcl-2 and A1 in vascular endothelial cells. *J. Biol. Chem.* **1998**, *273*, 13313–13316. [\[CrossRef\]](#) [\[PubMed\]](#)
139. Xiao, Z.; Liu, Q.; Mao, F.; Wu, J.; Lei, T. TNF- $\alpha$ -induced VEGF and MMP-9 expression promotes hemorrhagic transformation in pituitary adenomas. *Int. J. Mol. Sci.* **2011**, *12*, 4165–4179. [\[CrossRef\]](#) [\[PubMed\]](#)
140. Liang, Z.; Brooks, J.; Willard, M.; Liang, K.; Yoon, Y.; Kang, S.; Shim, H. CXCR4/CXCL12 axis promotes VEGF-mediated tumor angiogenesis through Akt signaling pathway. *Biochem. Biophys. Res. Commun.* **2007**, *359*, 716–722. [\[CrossRef\]](#) [\[PubMed\]](#)
141. Kryczek, I.; Lange, A.; Mottram, P.; Alvarez, X.; Cheng, P.; Hogan, M.; Moons, L.; Wei, S.; Zou, L.; Machelon, V.; et al. CXCL12 and vascular endothelial growth factor synergistically induce neoangiogenesis in human ovarian cancers. *Cancer Res.* **2005**, *65*, 465–472. [\[PubMed\]](#)
142. Motro, B.; Itin, A.; Sachs, L.; Keshet, E. Pattern of interleukin 6 gene expression in vivo suggests a role for this cytokine in angiogenesis. *Proc. Natl. Acad. Sci. USA* **1990**, *87*, 3092–3096. [\[CrossRef\]](#) [\[PubMed\]](#)
143. Nilsson, M.B.; Langley, R.R.; Fidler, I.J. Interleukin-6, secreted by human ovarian carcinoma cells, is a potent proangiogenic cytokine. *Cancer Res.* **2005**, *65*, 10794–10800. [\[CrossRef\]](#) [\[PubMed\]](#)
144. Rose-John, S.; Scheller, J.; Elson, G.; Jones, S.A. Interleukin-6 biology is coordinated by membrane-bound and soluble receptors: Role in inflammation and cancer. *J. Leukocyte Biol.* **2006**, *80*, 227–236. [\[CrossRef\]](#) [\[PubMed\]](#)
145. Hu, D.E.; Hori, Y.; Fan, T.P. Interleukin-8 stimulates angiogenesis in rats. *Inflammation* **1993**, *17*, 135–143. [\[CrossRef\]](#) [\[PubMed\]](#)
146. Koch, A.E.; Poverini, P.J.; Kunkel, S.L.; Harlow, L.A.; DiPietro, L.A.; Elner, V.M.; Elner, S.G.; Strieter, R.M. Interleukin-8 as a macrophage-derived mediator of angiogenesis. *Science* **1992**, *258*, 1798–1801. [\[CrossRef\]](#) [\[PubMed\]](#)
147. Stetler-Stevenson, W.G. Matrix metalloproteinases in angiogenesis: A moving target for therapeutic intervention. *J. Clin. Invest.* **1999**, *103*, 1237–1241. [\[CrossRef\]](#) [\[PubMed\]](#)
148. Lau, T.S.; Chan, L.K.; Wong, E.C.; Hui, C.W.; Sneddon, K.; Cheung, T.H.; Yim, S.F.; Lee, J.H.; Yeung, C.S.; Chung, T.K.; et al. A loop of cancer-stroma-cancer interaction promotes peritoneal metastasis of ovarian cancer via TNF $\alpha$ -TGF $\alpha$ -EGFR. *Oncogene* **2017**, *36*, 3576–3587. [\[CrossRef\]](#) [\[PubMed\]](#)
149. Scotton, C.J.; Wilson, J.L.; Scott, K.; Stamp, G.; Wilbanks, G.D.; Fricker, S.; Bridger, G.; Balkwill, F.R. Multiple actions of the chemokine CXCL12 on epithelial tumor cells in human ovarian cancer. *Cancer Res.* **2002**, *62*, 5930–5938. [\[PubMed\]](#)
150. Kulbe, H.; Hagemann, T.; Szlosarek, P.W.; Balkwill, F.R.; Wilson, J.L. The inflammatory cytokine tumor necrosis factor- $\alpha$  regulates chemokine receptor expression on ovarian cancer cells. *Cancer Res.* **2005**, *65*, 10355–10362. [\[CrossRef\]](#) [\[PubMed\]](#)
151. Kim, S.; Gwak, H.; Kim, H.S.; Kim, B.; Dhanasekaran, D.N.; Song, Y.S. Malignant ascites enhances migratory and invasive properties of ovarian cancer cells with membrane bound IL-6R in vitro. *Oncotarget* **2016**, *7*, 83148–83159. [\[CrossRef\]](#) [\[PubMed\]](#)
152. Wang, Y.; Niu, X.L.; Qu, Y.; Wu, J.; Zhu, Y.Q.; Sun, W.J.; Li, L.Z. Autocrine production of interleukin-6 confers cisplatin and paclitaxel resistance in ovarian cancer cells. *Cancer Lett.* **2010**, *295*, 110–123. [\[CrossRef\]](#) [\[PubMed\]](#)

153. Yung, M.M.; Tang, H.W.; Cai, P.C.; Leung, T.H.; Ngu, S.F.; Chan, K.K.; Xu, D.; Yang, H.; Ngan, H.Y.; Chan, D.W. GRO- $\alpha$  and IL-8 enhance ovarian cancer metastatic potential via the CXCR2-mediated TAK1/NF $\kappa$ B signaling cascade. *Theranostics* **2018**, *8*, 1270–1285. [[CrossRef](#)] [[PubMed](#)]
154. Cuello, M.; Ettenberg, S.A.; Nau, M.M.; Lipkowitz, S. Synergistic induction of apoptosis by the combination of trail and chemotherapy in chemoresistant ovarian cancer cells. *Gynecol. Oncol.* **2001**, *81*, 380–390. [[CrossRef](#)] [[PubMed](#)]
155. Song, Y.; Wu, J.; Oyesanya, R.A.; Lee, Z.; Mukherjee, A.; Fang, X. Sp-1 and c-Myc mediate lysophosphatidic acid-induced expression of vascular endothelial growth factor in ovarian cancer cells via a hypoxia-inducible factor-1-independent mechanism. *Clin. Cancer Res.* **2009**, *15*, 492–501. [[CrossRef](#)] [[PubMed](#)]
156. Pustilnik, T.B.; Estrella, V.; Wiener, J.R.; Mao, M.; Eder, A.; Watt, M.A.; Bast, R.C., Jr.; Mills, G.B. Lysophosphatidic acid induces urokinase secretion by ovarian cancer cells. *Clin. Cancer Res.* **1999**, *5*, 3704–3710. [[PubMed](#)]
157. Duffy, M.J. Proteases as prognostic markers in cancer. *Clin. Cancer Res.* **1996**, *2*, 613–618. [[PubMed](#)]
158. Masferrer, J.L.; Leahy, K.M.; Koki, A.T.; Zweifel, B.S.; Settle, S.L.; Woerner, B.M.; Edwards, D.A.; Flickinger, A.G.; Moore, R.J.; Seibert, K. Antiangiogenic and antitumor activities of cyclooxygenase-2 inhibitors. *Cancer Res.* **2000**, *60*, 1306–1311. [[PubMed](#)]
159. Liu, X.H.; Yao, S.; Kirschenbaum, A.; Levine, A.C. NS398, a selective cyclooxygenase-2 inhibitor, induces apoptosis and down-regulates bcl-2 expression in LNCaP cells. *Cancer Res.* **1998**, *58*, 4245–4249. [[PubMed](#)]
160. Yeung, T.L.; Leung, C.S.; Wong, K.K.; Samimi, G.; Thompson, M.S.; Liu, J.; Zaid, T.M.; Ghosh, S.; Birrer, M.J.; Mok, S.C. TGF- $\beta$  modulates ovarian cancer invasion by upregulating CAF-derived versican in the tumor microenvironment. *Cancer Res.* **2013**, *73*, 5016–5028. [[CrossRef](#)] [[PubMed](#)]
161. Frankel, A.; Mills, G.B. Peptide and lipid growth factors decrease cis-diamminedichloroplatinum-induced cell death in human ovarian cancer cells. *Clin. Cancer Res.* **1996**, *2*, 1307–1313. [[PubMed](#)]
162. Gao, Y.; Shan, N.; Zhao, C.; Wang, Y.; Xu, F.; Li, J.; Yu, X.; Gao, L.; Yi, Z. LY2109761 enhances cisplatin antitumor activity in ovarian cancer cells. *Int J. Clin. Exp. Pathol.* **2015**, *8*, 4923–4932. [[PubMed](#)]
163. Hu, Y.L.; Tee, M.K.; Goetzl, E.J.; Auersperg, N.; Mills, G.B.; Ferrara, N.; Jaffe, R.B. Lysophosphatidic acid induction of vascular endothelial growth factor expression in human ovarian cancer cells. *J. Natl. Cancer Inst.* **2001**, *93*, 762–768. [[CrossRef](#)] [[PubMed](#)]
164. Forsythe, J.A.; Jiang, B.H.; Iyer, N.V.; Agani, F.; Leung, S.W.; Koos, R.D.; Semenza, G.L. Activation of vascular endothelial growth factor gene transcription by hypoxia-inducible factor 1. *Mol. Cell. Biol.* **1996**, *16*, 4604–4613. [[CrossRef](#)] [[PubMed](#)]
165. Tan, D.S.; Agarwal, R.; Kaye, S.B. Mechanisms of transcoelomic metastasis in ovarian cancer. *Lancet Oncol.* **2006**, *7*, 925–934. [[CrossRef](#)]
166. Fidler, I.J. The pathogenesis of cancer metastasis: The ‘seed and soil’ hypothesis revisited. *Nat. Rev. Cancer* **2003**, *3*, 453–458. [[CrossRef](#)] [[PubMed](#)]
167. Gupta, G.P.; Massague, J. Cancer metastasis: Building a framework. *Cell* **2006**, *127*, 679–695. [[CrossRef](#)] [[PubMed](#)]
168. Talmadge, J.E.; Fidler, I.J. AACR centennial series: The biology of cancer metastasis: Historical perspective. *Cancer Res.* **2010**, *70*, 5649–5669. [[CrossRef](#)] [[PubMed](#)]
169. Yang-Hartwich, Y.; Gurea-Soteras, M.; Sumi, N.; Joo, W.D.; Holmberg, J.C.; Craveiro, V.; Alvero, A.B.; Mor, G. Ovulation and extra-ovarian origin of ovarian cancer. *Sci. Rep.* **2014**, *4*, 6116. [[CrossRef](#)] [[PubMed](#)]
170. Russo, A.; Czarnecki, A.A.; Dean, M.; Modi, D.A.; Lantvit, D.D.; Hardy, L.; Baligod, S.; Davis, D.A.; Wei, J.J.; Burdette, J.E. PTEN loss in the fallopian tube induces hyperplasia and ovarian tumor formation. *Oncogene* **2018**, *37*, 1976–1990. [[CrossRef](#)] [[PubMed](#)]
171. Cain, R.J.; Ridley, A.J. Phosphoinositide 3-kinases in cell migration. *Biol. Cell* **2009**, *101*, 13–29. [[CrossRef](#)] [[PubMed](#)]
172. Raftopoulos, M.; Hall, A. Cell migration: Rho GTPases lead the way. *Dev. Biol.* **2004**, *265*, 23–32. [[CrossRef](#)] [[PubMed](#)]
173. Devreotes, P.; Horwitz, A.R. Signaling networks that regulate cell migration. *Cold Spring Harb Perspect. Biol.* **2015**, *7*, a005959. [[CrossRef](#)] [[PubMed](#)]
174. Robinson-Smith, T.M.; Isaacsohn, I.; Mercer, C.A.; Zhou, M.; Van Rooijen, N.; Hussein, N.; McFarland-Mancini, M.M.; Drew, A.F. Macrophages mediate inflammation-enhanced metastasis of ovarian tumors in mice. *Cancer Res.* **2007**, *67*, 5708–5716. [[CrossRef](#)] [[PubMed](#)]

175. Wilkosz, S.; Ireland, G.; Khwaja, N.; Walker, M.; Butt, R.; de Giorgio-Miller, A.; Herrick, S.E. A comparative study of the structure of human and murine greater omentum. *Anat. Embryol.* **2005**, *209*, 251–261. [[CrossRef](#)] [[PubMed](#)]
176. Tang, J.; Pulliam, N.; Ozes, A.; Buechlein, A.; Ding, N.; Keer, H.; Rusch, D.; O'Hagan, H.; Stack, M.S.; Nephew, K.P. Epigenetic Targeting of Adipocytes Inhibits High-Grade Serous Ovarian Cancer Cell Migration and Invasion. *Mol. Cancer Res.* **2018**. [[CrossRef](#)] [[PubMed](#)]
177. Meza-Perez, S.; Randall, T.D. Immunological Functions of the Omentum. *Trends Immunol.* **2017**, *38*, 526–536. [[CrossRef](#)] [[PubMed](#)]
178. Gerber, S.A.; Rybalko, V.Y.; Bigelow, C.E.; Lugade, A.A.; Foster, T.H.; Frelinger, J.G.; Lord, E.M. Preferential attachment of peritoneal tumor metastases to omental immune aggregates and possible role of a unique vascular microenvironment in metastatic survival and growth. *Am. J. Pathol.* **2006**, *169*, 1739–1752. [[CrossRef](#)] [[PubMed](#)]
179. Clark, R.; Krishnan, V.; Schoof, M.; Rodriguez, I.; Theriault, B.; Chekmareva, M.; Rinker-Schaeffer, C. Milky spots promote ovarian cancer metastatic colonization of peritoneal adipose in experimental models. *Am. J. Pathol.* **2013**, *183*, 576–591. [[CrossRef](#)] [[PubMed](#)]
180. Szatrowski, T.P.; Nathan, C.F. Production of large amounts of hydrogen peroxide by human tumor cells. *Cancer Res.* **1991**, *51*, 794–798. [[PubMed](#)]
181. Wang, Y.; Ma, J.; Shen, H.; Wang, C.; Sun, Y.; Howell, S.B.; Lin, X. Reactive oxygen species promote ovarian cancer progression via the HIF-1 $\alpha$ /LOX/E-cadherin pathway. *Oncol. Rep.* **2014**, *32*, 2150–2158. [[CrossRef](#)] [[PubMed](#)]
182. Kundu, N.; Zhang, S.; Fulton, A.M. Sublethal oxidative stress inhibits tumor cell adhesion and enhances experimental metastasis of murine mammary carcinoma. *Clin. Exp. Metast.* **1995**, *13*, 16–22. [[CrossRef](#)]
183. Haskill, S.; Becker, S.; Fowler, W.; Walton, L. Mononuclear-cell infiltration in ovarian cancer. I. Inflammatory-cell infiltrates from tumour and ascites material. *Br. J. Cancer* **1982**, *45*, 728–736. [[CrossRef](#)] [[PubMed](#)]
184. Wang, J.M.; Griffin, J.D.; Rambaldi, A.; Chen, Z.G.; Mantovani, A. Induction of monocyte migration by recombinant macrophage colony-stimulating factor. *J. Immunol.* **1988**, *141*, 575–579. [[PubMed](#)]
185. Ramakrishnan, S.; Xu, F.J.; Brandt, S.J.; Niedel, J.E.; Bast, R.C., Jr.; Brown, E.L. Constitutive production of macrophage colony-stimulating factor by human ovarian and breast cancer cell lines. *J. Clin. Investig.* **1989**, *83*, 921–926. [[CrossRef](#)] [[PubMed](#)]
186. Plante, M.; Rubin, S.C.; Wong, G.Y.; Federici, M.G.; Finstad, C.L.; Gastl, G.A. Interleukin-6 level in serum and ascites as a prognostic factor in patients with epithelial ovarian cancer. *Cancer* **1994**, *73*, 1882–1888. [[CrossRef](#)]
187. Scambia, G.; Testa, U.; Panici, P.B.; Martucci, R.; Foti, E.; Petrini, M.; Amoroso, M.; Masciullo, V.; Peschle, C.; Mancuso, S. Interleukin-6 serum levels in patients with gynecological tumors. *Int. J. Cancer* **1994**, *57*, 318–323. [[CrossRef](#)] [[PubMed](#)]
188. Tempfer, C.; Zeisler, H.; Sliutz, G.; Haeusler, G.; Hanzal, E.; Kainz, C. Serum evaluation of interleukin 6 in ovarian cancer patients. *Gynecol. Oncol.* **1997**, *66*, 27–30. [[CrossRef](#)] [[PubMed](#)]
189. Isobe, A.; Sawada, K.; Kinose, Y.; Ohyagi-Hara, C.; Nakatsuka, E.; Makino, H.; Ogura, T.; Mizuno, T.; Suzuki, N.; Morii, E.; et al. Interleukin 6 receptor is an independent prognostic factor and a potential therapeutic target of ovarian cancer. *PLoS ONE* **2015**, *10*, e0118080. [[CrossRef](#)] [[PubMed](#)]
190. Fishman, D.A.; Liu, Y.; Ellerbroek, S.M.; Stack, M.S. Lysophosphatidic acid promotes matrix metalloproteinase (MMP) activation and MMP-dependent invasion in ovarian cancer cells. *Cancer Res.* **2001**, *61*, 3194–3199. [[PubMed](#)]
191. Reiser, C.O.; Lanz, T.; Hofmann, F.; Hofer, G.; Rupprecht, H.D.; Goppelt-Struebe, M. Lysophosphatidic acid-mediated signal-transduction pathways involved in the induction of the early-response genes prostaglandin G/H synthase-2 and Egr-1: A critical role for the mitogen-activated protein kinase p38 and for Rho proteins. *Biochem. J.* **1998**, *330* (Pt 3), 1107–1114. [[CrossRef](#)] [[PubMed](#)]
192. Moolenaar, W.H.; Kruijer, W.; Tilly, B.C.; Verlaan, I.; Bierman, A.J.; de Laat, S.W. Growth factor-like action of phosphatidic acid. *Nature* **1986**, *323*, 171–173. [[CrossRef](#)] [[PubMed](#)]
193. Schwartz, B.M.; Hong, G.; Morrison, B.H.; Wu, W.; Baudhuin, L.M.; Xiao, Y.J.; Mok, S.C.; Xu, Y. Lysophospholipids increase interleukin-8 expression in ovarian cancer cells. *Gynecol. Oncol.* **2001**, *81*, 291–300. [[CrossRef](#)] [[PubMed](#)]



194. Yu, X.; Zhang, Y.; Chen, H. LPA receptor 1 mediates LPA-induced ovarian cancer metastasis: An in vitro and in vivo study. *BMC Cancer* **2016**, *16*, 846. [[CrossRef](#)] [[PubMed](#)]
195. Loizzi, V.; Del Vecchio, V.; Gargano, G.; De Liso, M.; Kardashi, A.; Naglieri, E.; Resta, L.; Cicinelli, E.; Cormio, G. Biological Pathways Involved in Tumor Angiogenesis and Bevacizumab Based Anti-Angiogenic Therapy with Special References to Ovarian Cancer. *Int. J. Mol. Sci.* **2017**, *18*, 1967. [[CrossRef](#)] [[PubMed](#)]
196. Le Page, C.; Puiffe, M.L.; Meunier, L.; Zietarska, M.; de Ladurantaye, M.; Tonin, P.N.; Provencher, D.; Mes-Masson, A.M. BMP-2 signaling in ovarian cancer and its association with poor prognosis. *J. Ovarian Res.* **2009**, *2*, 4. [[CrossRef](#)] [[PubMed](#)]
197. Ozols, R.F. Treatment goals in ovarian cancer. *Int. J. Gynecol. Cancer* **2005**, *15* (Suppl. 1), 3–11. [[CrossRef](#)] [[PubMed](#)]
198. Koti, M.; Siu, A.; Clement, I.; Bidarimath, M.; Turashvili, G.; Edwards, A.; Rahimi, K.; Mes-Masson, A.M.; Squire, J.A. A distinct pre-existing inflammatory tumour microenvironment is associated with chemotherapy resistance in high-grade serous epithelial ovarian cancer. *Br. J. Cancer* **2015**, *112*, 1215–1222. [[CrossRef](#)] [[PubMed](#)]
199. Stewart, D.J. Mechanisms of resistance to cisplatin and carboplatin. *Crit. Rev. Oncol. Hematol.* **2007**, *63*, 12–31. [[CrossRef](#)] [[PubMed](#)]
200. Damiano, J.S.; Cress, A.E.; Hazlehurst, L.A.; Shtil, A.A.; Dalton, W.S. Cell adhesion mediated drug resistance (CAM-DR): Role of integrins and resistance to apoptosis in human myeloma cell lines. *Blood* **1999**, *93*, 1658–1667. [[PubMed](#)]
201. Meads, M.B.; Hazlehurst, L.A.; Dalton, W.S. The bone marrow microenvironment as a tumor sanctuary and contributor to drug resistance. *Clin. Cancer Res.* **2008**, *14*, 2519–2526. [[CrossRef](#)] [[PubMed](#)]
202. Catlett-Falcone, R.; Landowski, T.H.; Oshiro, M.M.; Turkson, J.; Levitzki, A.; Savino, R.; Ciliberto, G.; Moscinski, L.; Fernandez-Luna, J.L.; Nunez, G.; et al. Constitutive activation of Stat3 signaling confers resistance to apoptosis in human U266 myeloma cells. *Immunity* **1999**, *10*, 105–115. [[CrossRef](#)]
203. Frassanito, M.A.; Cusmai, A.; Iodice, G.; Dammacco, F. Autocrine interleukin-6 production and highly malignant multiple myeloma: Relation with resistance to drug-induced apoptosis. *Blood* **2001**, *97*, 483–489. [[CrossRef](#)] [[PubMed](#)]
204. Meads, M.B.; Gatenby, R.A.; Dalton, W.S. Environment-mediated drug resistance: A major contributor to minimal residual disease. *Nat. Rev. Cancer* **2009**, *9*, 665–674. [[CrossRef](#)] [[PubMed](#)]
205. Cohen, S.; Mehrabi, S.; Yao, X.; Millingen, S.; Aikionbare, F.O. Reactive Oxygen Species and Serous Epithelial Ovarian Adenocarcinoma. *Cancer Res. J.* **2016**, *4*, 106–114. [[CrossRef](#)] [[PubMed](#)]
206. St-Pierre, J.; Drori, S.; Uldry, M.; Silvaggi, J.M.; Rhee, J.; Jager, S.; Handschin, C.; Zheng, K.; Lin, J.; Yang, W.; et al. Suppression of reactive oxygen species and neurodegeneration by the PGC-1 transcriptional coactivators. *Cell* **2006**, *127*, 397–408. [[CrossRef](#)] [[PubMed](#)]
207. Kim, B.; Jung, J.W.; Jung, J.; Han, Y.; Suh, D.H.; Kim, H.S.; Dhanasekaran, D.N.; Song, Y.S. PGC1 $\alpha$  induced by reactive oxygen species contributes to chemoresistance of ovarian cancer cells. *Oncotarget* **2017**, *8*, 60299–60311. [[PubMed](#)]
208. Duan, Z.; Feller, A.J.; Penson, R.T.; Chabner, B.A.; Seiden, M.V. Discovery of differentially expressed genes associated with paclitaxel resistance using cDNA array technology: Analysis of interleukin (IL) 6, IL-8, and monocyte chemotactic protein 1 in the paclitaxel-resistant phenotype. *Clin. Cancer Res.* **1999**, *5*, 3445–3453. [[PubMed](#)]
209. Duan, Z.; Foster, R.; Bell, D.A.; Mahoney, J.; Wolak, K.; Vaidya, A.; Hampel, C.; Lee, H.; Seiden, M.V. Signal transducers and activators of transcription 3 pathway activation in drug-resistant ovarian cancer. *Clin. Cancer Res.* **2006**, *12*, 5055–5063. [[CrossRef](#)] [[PubMed](#)]
210. Abdollahi, T.; Robertson, N.M.; Abdollahi, A.; Litwack, G. Identification of interleukin 8 as an inhibitor of tumor necrosis factor-related apoptosis-inducing ligand-induced apoptosis in the ovarian carcinoma cell line OVCAR3. *Cancer Res.* **2003**, *63*, 4521–4526. [[PubMed](#)]
211. Wiley, S.R.; Schooley, K.; Smolak, P.J.; Din, W.S.; Huang, C.P.; Nicholl, J.K.; Sutherland, G.R.; Smith, T.D.; Rauch, C.; Smith, C.A.; et al. Identification and characterization of a new member of the TNF family that induces apoptosis. *Immunity* **1995**, *3*, 673–682. [[CrossRef](#)]
212. Walczak, H.; Miller, R.E.; Ariail, K.; Gliniak, B.; Griffith, T.S.; Kubin, M.; Chin, W.; Jones, J.; Woodward, A.; Le, T.; et al. Tumoricidal activity of tumor necrosis factor-related apoptosis-inducing ligand in vivo. *Nat. Med.* **1999**, *5*, 157–163. [[CrossRef](#)] [[PubMed](#)]



213. Bristow, R.E.; Baldwin, R.L.; Yamada, S.D.; Korc, M.; Karlan, B.Y. Altered expression of transforming growth factor-beta ligands and receptors in primary and recurrent ovarian carcinoma. *Cancer* **1999**, *85*, 658–668. [[CrossRef](#)]
214. Li, M.; Balch, C.; Montgomery, J.S.; Jeong, M.; Chung, J.H.; Yan, P.; Huang, T.H.; Kim, S.; Nephew, K.P. Integrated analysis of DNA methylation and gene expression reveals specific signaling pathways associated with platinum resistance in ovarian cancer. *BMC Med. Genom.* **2009**, *2*, 34. [[CrossRef](#)] [[PubMed](#)]
215. Psyrri, A.; Kassab, M.; Yu, Z.; Bamias, A.; Weinberger, P.M.; Markakis, S.; Kowalski, D.; Camp, R.L.; Rimm, D.L.; Dimopoulos, M.A. Effect of epidermal growth factor receptor expression level on survival in patients with epithelial ovarian cancer. *Clin. Cancer Res.* **2005**, *11*, 8637–8643. [[CrossRef](#)] [[PubMed](#)]
216. Crew, A.J.; Langdon, S.P.; Miller, E.P.; Miller, W.R. Mitogenic effects of epidermal growth factor and transforming growth factor- $\alpha$  on EGF-receptor positive human ovarian carcinoma cell lines. *Eur. J. Cancer* **1992**, *28*, 337–341. [[CrossRef](#)]
217. Ferrandina, G.; Ranelletti, F.O.; Martinelli, E.; Paglia, A.; Zannoni, G.F.; Scambia, G. Cyclo-oxygenase-2 (Cox-2) expression and resistance to platinum versus platinum/paclitaxel containing chemotherapy in advanced ovarian cancer. *BMC Cancer* **2006**, *6*, 182. [[CrossRef](#)] [[PubMed](#)]
218. Ferrandina, G.; Lauriola, L.; Zannoni, G.F.; Fagotti, A.; Fanfani, F.; Legge, F.; Maggiano, N.; Gessi, M.; Mancuso, S.; Ranelletti, F.O.; et al. Increased cyclooxygenase-2 (COX-2) expression is associated with chemotherapy resistance and outcome in ovarian cancer patients. *Ann. Oncol.* **2002**, *13*, 1205–1211. [[CrossRef](#)] [[PubMed](#)]
219. Kim, A.; Ueda, Y.; Naka, T.; Enomoto, T. Therapeutic strategies in epithelial ovarian cancer. *J. Exp. Clin. Cancer Res.* **2012**, *31*, 14. [[CrossRef](#)] [[PubMed](#)]
220. Ellis, L.M.; Hicklin, D.J. VEGF-targeted therapy: Mechanisms of anti-tumour activity. *Nat. Rev. Cancer* **2008**, *8*, 579–591. [[CrossRef](#)] [[PubMed](#)]
221. Aghajanian, C.; Blank, S.V.; Goff, B.A.; Judson, P.L.; Teneriello, M.G.; Husain, A.; Sovak, M.A.; Yi, J.; Nycum, L.R. OCEANS: A randomized, double-blind, placebo-controlled phase III trial of chemotherapy with or without bevacizumab in patients with platinum-sensitive recurrent epithelial ovarian, primary peritoneal, or fallopian tube cancer. *J. Clin. Oncol.* **2012**, *30*, 2039–2045. [[CrossRef](#)] [[PubMed](#)]
222. Burger, R.A.; Brady, M.F.; Bookman, M.A.; Fleming, G.F.; Monk, B.J.; Huang, H.; Mannel, R.S.; Homesley, H.D.; Fowler, J.; Greer, B.E.; et al. Incorporation of bevacizumab in the primary treatment of ovarian cancer. *N. Engl. J. Med.* **2011**, *365*, 2473–2483. [[CrossRef](#)] [[PubMed](#)]
223. Perren, T.J.; Swart, A.M.; Pfisterer, J.; Ledermann, J.A.; Pujade-Lauraine, E.; Kristensen, G.; Carey, M.S.; Beale, P.; Cervantes, A.; Kurzeder, C.; et al. A phase 3 trial of bevacizumab in ovarian cancer. *N. Engl. J. Med.* **2011**, *365*, 2484–2496. [[CrossRef](#)] [[PubMed](#)]
224. Brown, E.R.; Charles, K.A.; Hoare, S.A.; Rye, R.L.; Jodrell, D.I.; Aird, R.E.; Vora, R.; Prabhakar, U.; Nakada, M.; Corringham, R.E.; et al. A clinical study assessing the tolerability and biological effects of infliximab, a TNF- $\alpha$  inhibitor, in patients with advanced cancer. *Ann. Oncol.* **2008**, *19*, 1340–1346. [[CrossRef](#)] [[PubMed](#)]
225. Guo, Y.; Nemeth, J.; O'Brien, C.; Susa, M.; Liu, X.; Zhang, Z.; Choy, E.; Mankin, H.; Hornicek, F.; Duan, Z. Effects of siltuximab on the IL-6-induced signaling pathway in ovarian cancer. *Clin. Cancer Res.* **2010**, *16*, 5759–5769. [[CrossRef](#)] [[PubMed](#)]
226. Xu, S.; Grande, F.; Garofalo, A.; Neamati, N. Discovery of a novel orally active small-molecule gp130 inhibitor for the treatment of ovarian cancer. *Mol. Cancer Ther.* **2013**, *12*, 937–949. [[CrossRef](#)] [[PubMed](#)]
227. Dijkgraaf, E.M.; Santegoets, S.J.; Reyners, A.K.; Goedemans, R.; Wouters, M.C.; Kenter, G.G.; van Erkel, A.R.; van Poelgeest, M.I.; Nijman, H.W.; van der Hoeven, J.J.; et al. A phase I trial combining carboplatin/doxorubicin with tocilizumab, an anti-IL-6R monoclonal antibody, and interferon- $\alpha$ 2b in patients with recurrent epithelial ovarian cancer. *Ann. Oncol.* **2015**, *26*, 2141–2149. [[CrossRef](#)] [[PubMed](#)]
228. Bhola, N.E.; Balko, J.M.; Dugger, T.C.; Kuba, M.G.; Sanchez, V.; Sanders, M.; Stanford, J.; Cook, R.S.; Arteaga, C.L. TGF-beta inhibition enhances chemotherapy action against triple-negative breast cancer. *J. Clin. Invest.* **2013**, *123*, 1348–1358. [[CrossRef](#)] [[PubMed](#)]
229. Alsina-Sanchis, E.; Figueras, A.; Lahiguera, A.; Gil-Martin, M.; Pardo, B.; Piulats, J.M.; Marti, L.; Ponce, J.; Matias-Guiu, X.; Vidal, A.; et al. TGFbeta Controls Ovarian Cancer Cell Proliferation. *Int. J. Mol. Sci.* **2017**, *18*, 1658. [[CrossRef](#)] [[PubMed](#)]

230. Secord, A.A.; Blessing, J.A.; Armstrong, D.K.; Rodgers, W.H.; Miner, Z.; Barnes, M.N.; Lewandowski, G.; Mannel, R.S.; Gynecologic Oncology, G. Phase II trial of cetuximab and carboplatin in relapsed platinum-sensitive ovarian cancer and evaluation of epidermal growth factor receptor expression: A Gynecologic Oncology Group study. *Gynecol. Oncol.* **2008**, *108*, 493–499. [[CrossRef](#)] [[PubMed](#)]
231. Chekerov, R.; Klare, P.; Krabisch, P.; Potenberg, J.; Heinrich, G.; Mueller, L.; Kurbacher, C.M.; Grischke, E.-M.; Braicu, E.I.; Wimberger, P.; et al. Panitumumab in platinum-sensitive epithelial ovarian cancer patients with KRAS wild-type: The PROVE-study, a phase II randomized multicenter study of the North-Eastern German Society of Gynaecologic Oncology. *J. Clin. Oncol.* **2017**, *35* (Suppl. 1), 5558.



© 2018 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).